

# An overview of genomics and sequencing terminology and practices

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Computational Biology and  
Genomics Workshop

Todos Santos Center  
April 9-13, 2018



There is a glossary available online that explains many of these terms

- Transcriptome
- Variant
- WGS

### 16S

The **16S** ribosomal RNA gene is present in all bacterial and archaeal genomes. This gene is sufficiently conserved that primers that anneal to conserved regions of the gene will amplify essentially any prokaryotic 16S rRNA gene. These PCR products (amplicons) can be sequenced to provide a survey of microbial diversity in a sample.

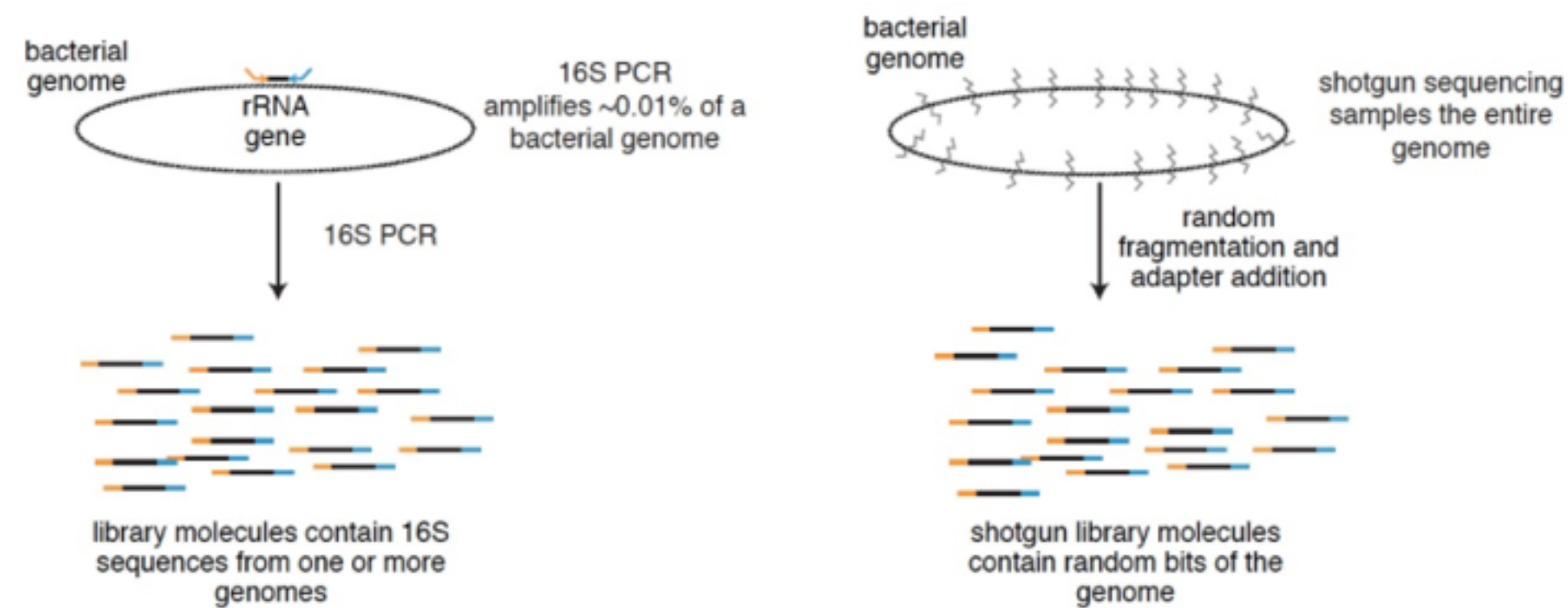


Figure: 16S vs. shotgun sequencing.

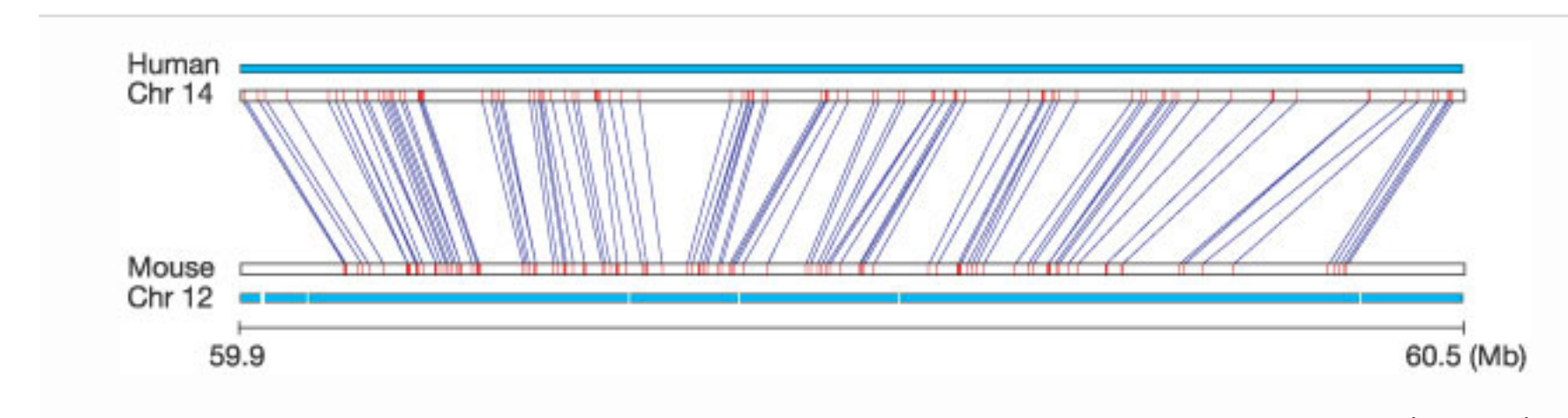
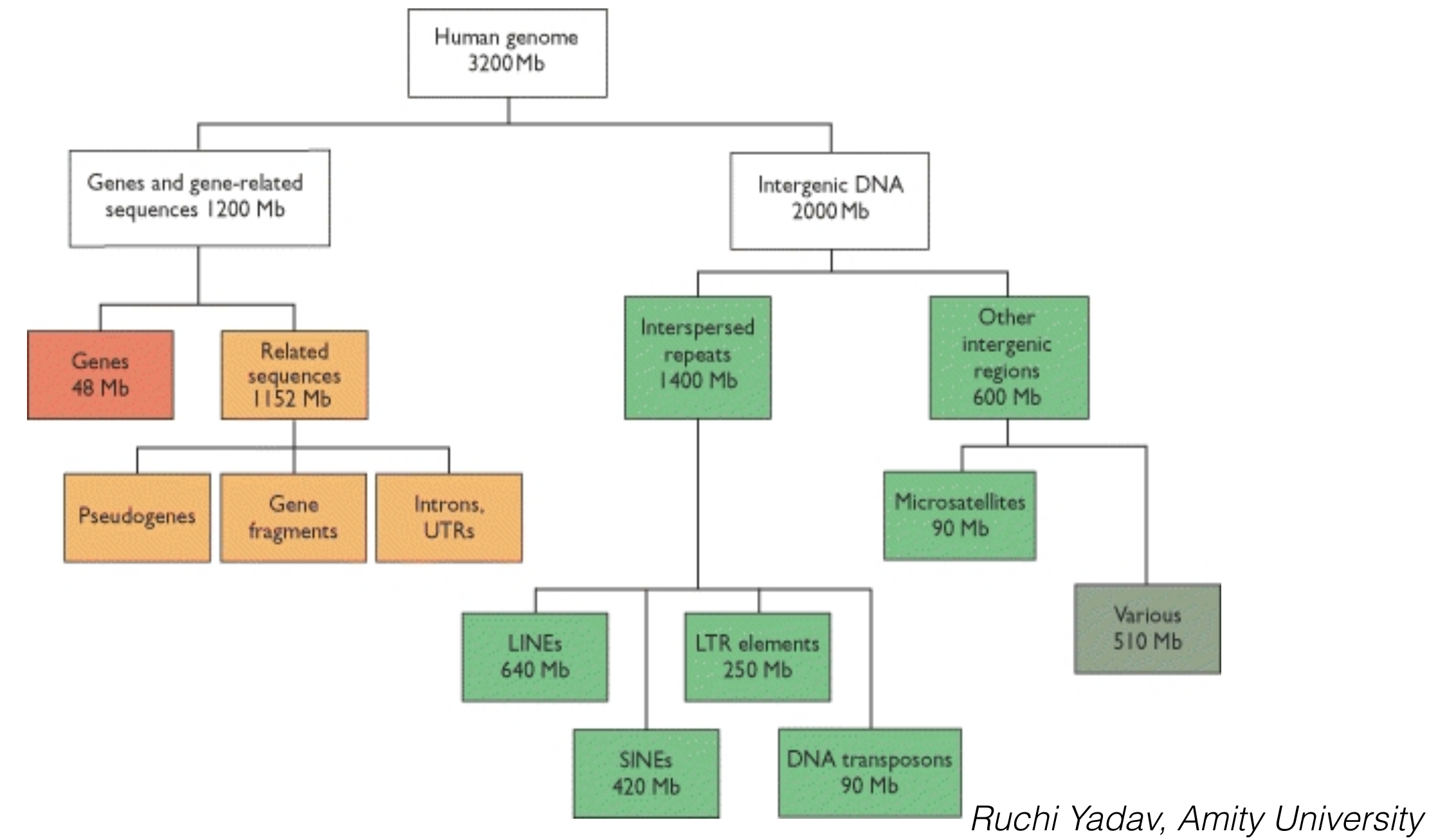
### Adapter

Most NGS instruments require that dsDNA of known sequence be added to the 2 ends of library molecules that will be sequenced on the instrument. Adapters can be added in a variety of ways to starting nucleic acid molecules during library



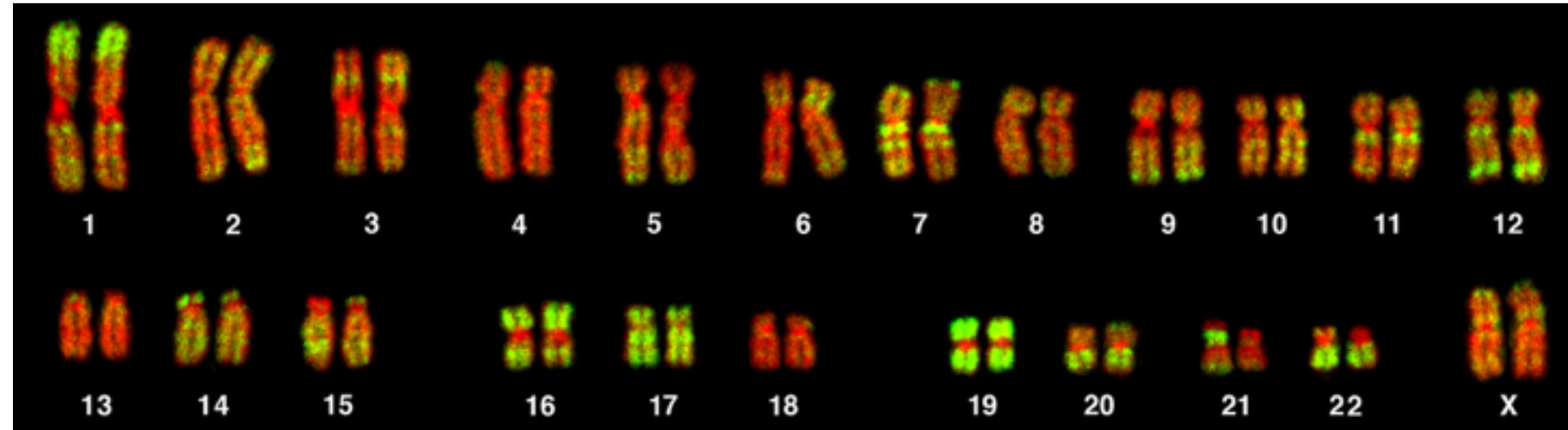
# Genomics is the study of any of a number of attributes of genome or genomes

- Genome:
  - size
  - sequence
  - structure / variation
  - evolution
- Gene:
  - structure
  - expression
- Comparative genomics
- Epigenomics
- Metagenomics
- Transcriptomics
- Other -omics: Proteomics/Metabolomics



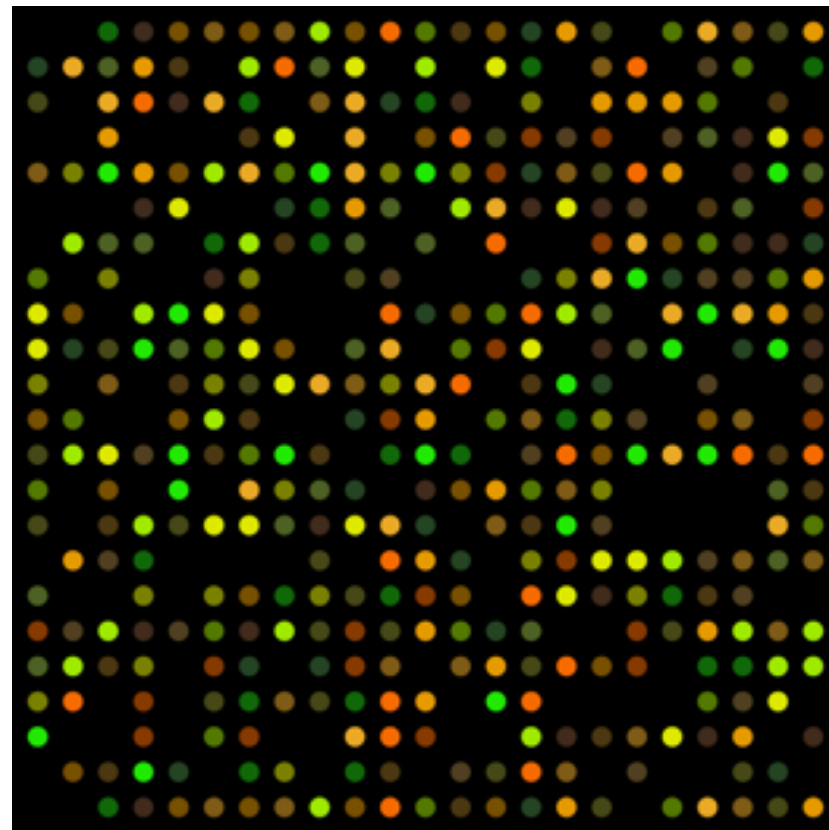
Nature (2002) Mouse Genome

# Genomics isn't the same thing as sequencing, but they're increasingly related

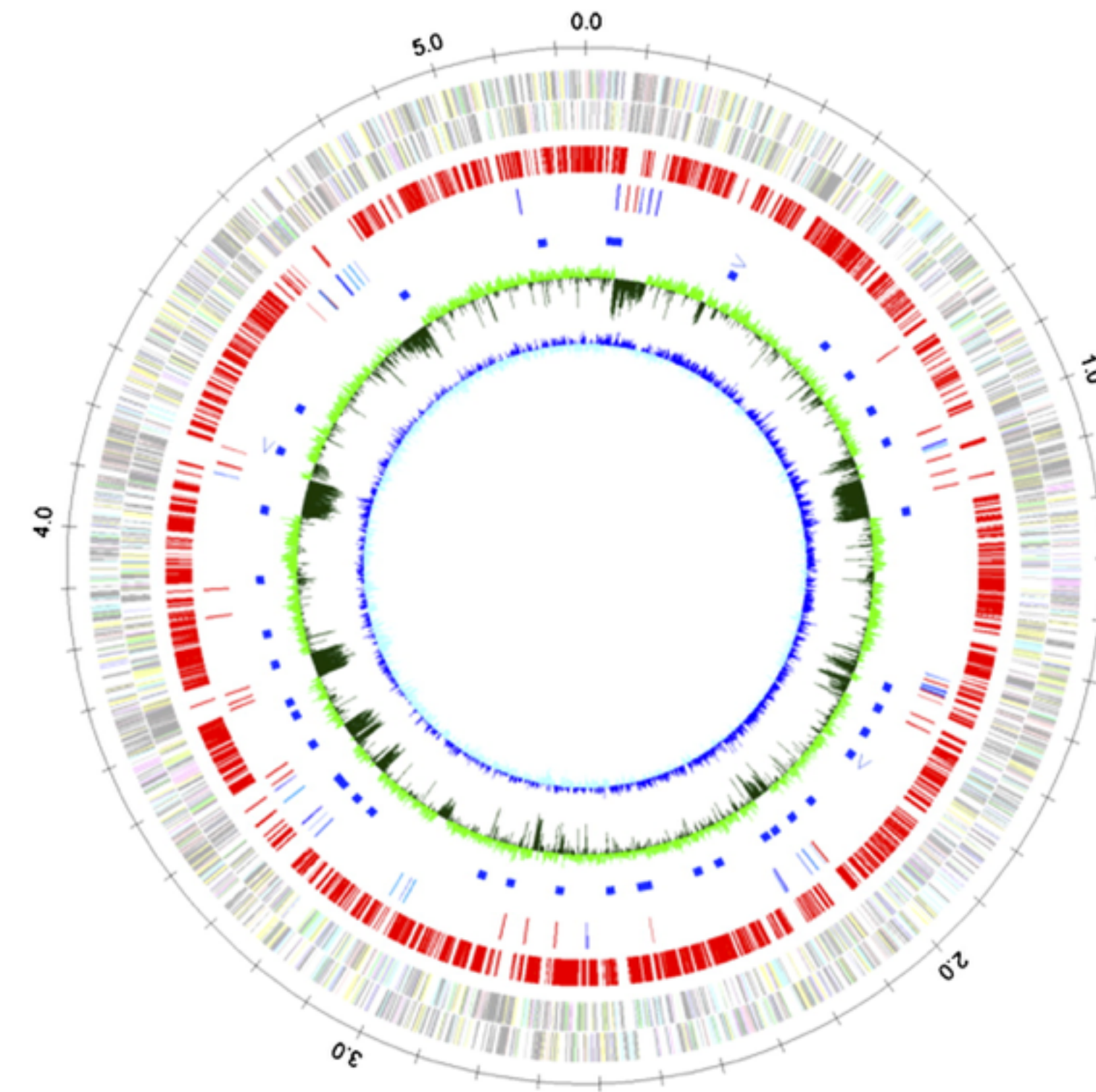


*Bolzer et al (2005) PLoS Biol*

Microarray



*Wikimedia commons*

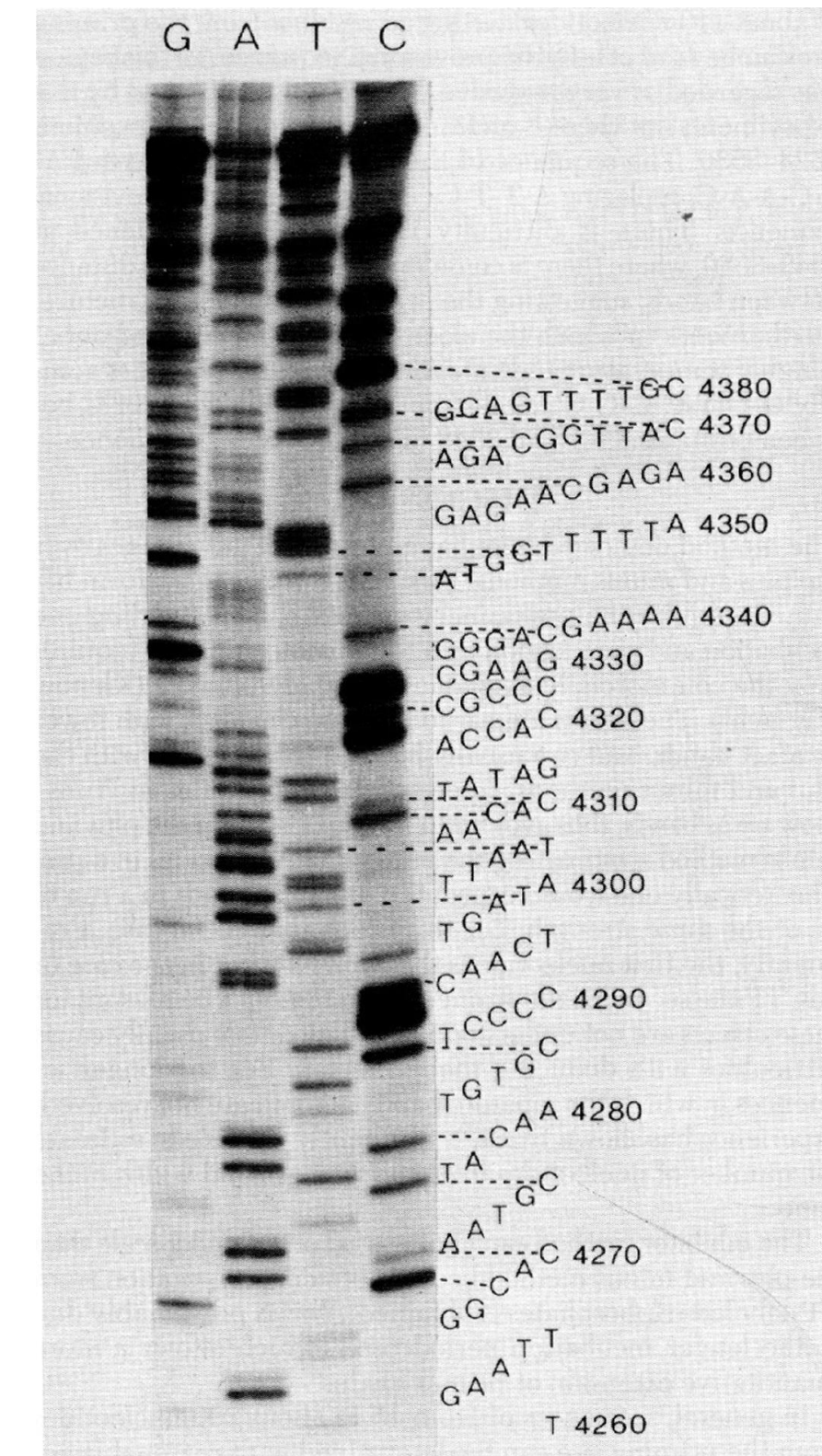
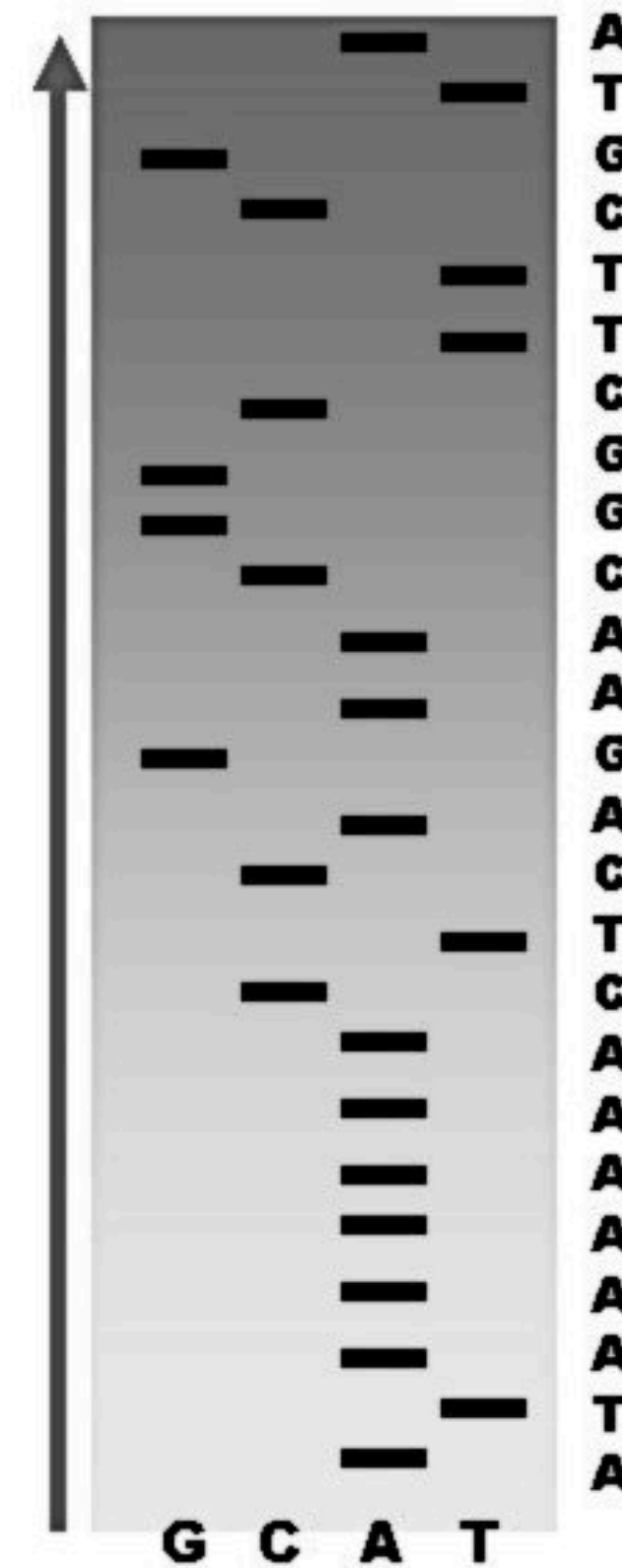
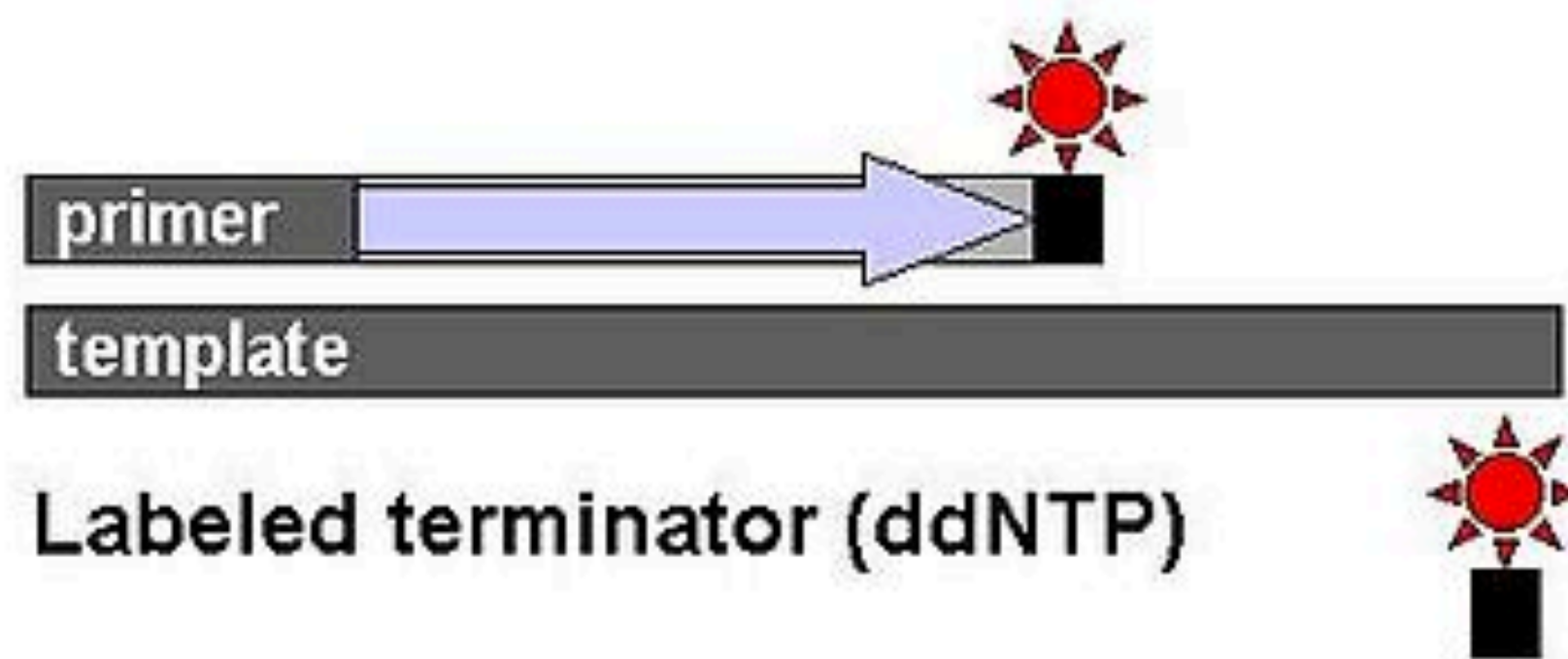
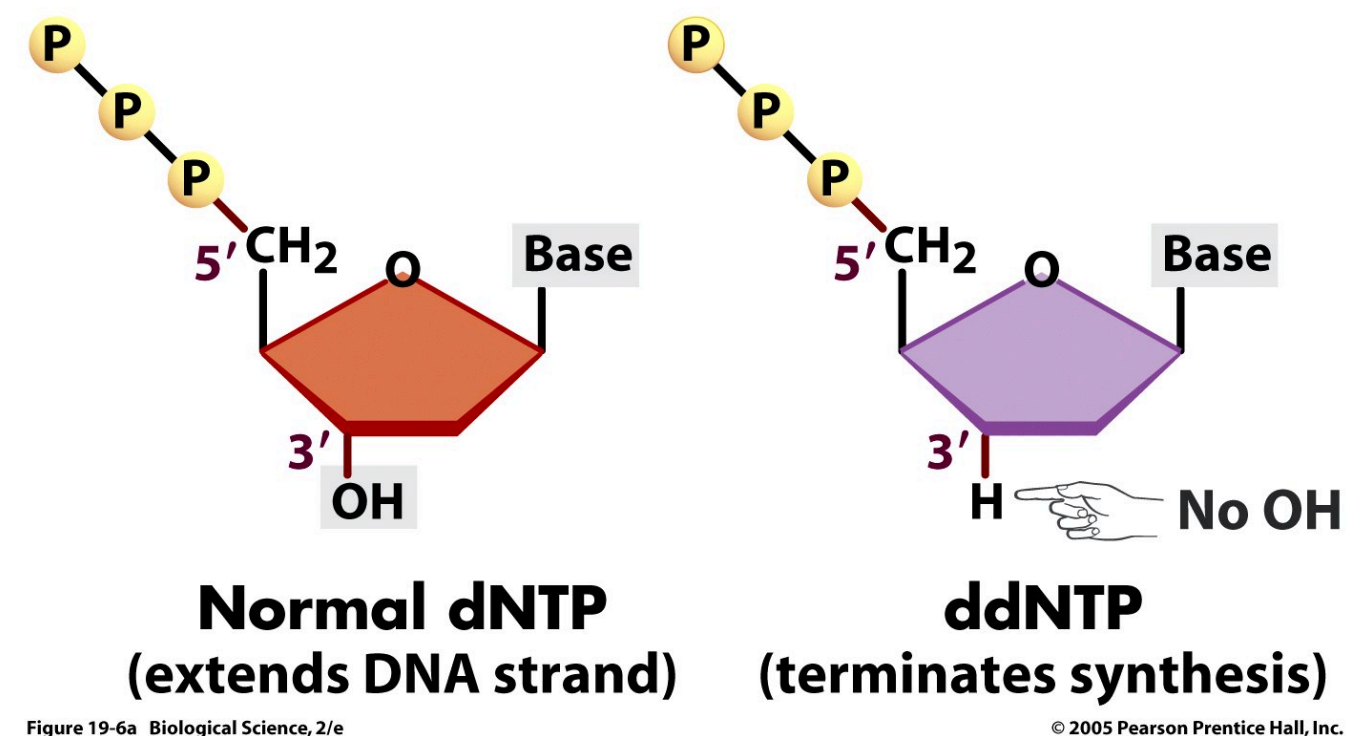


*Nakazawa et al (2009) Genome Research*



# Sanger Sequencing (1977): sequencing 1 target at a time

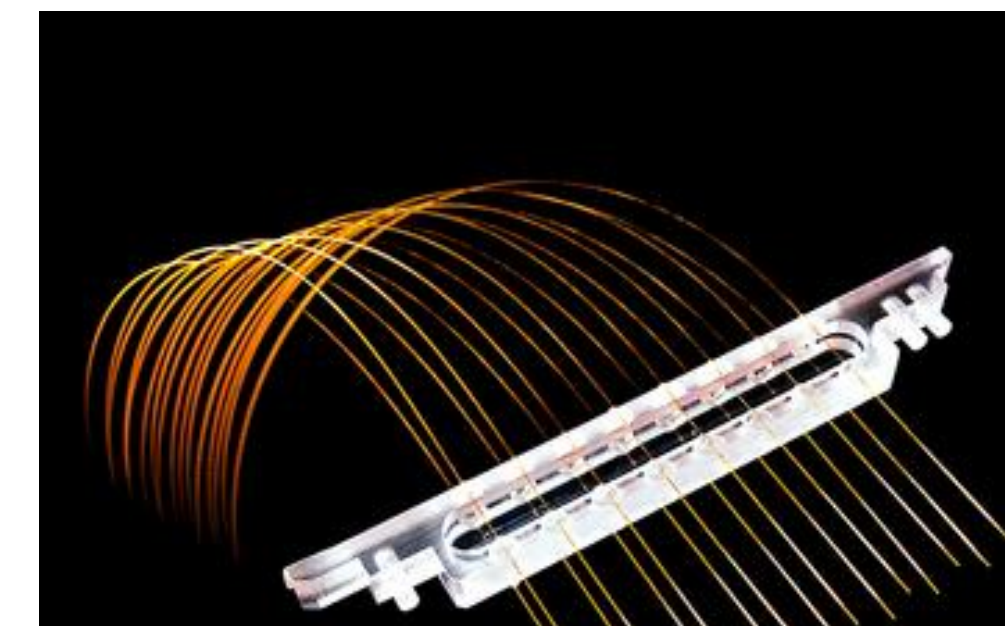
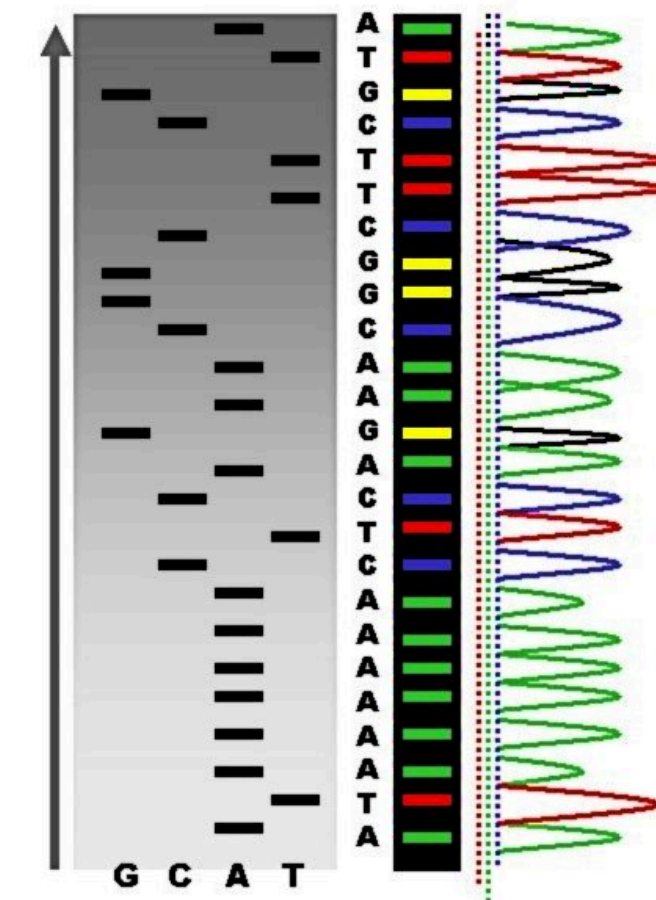
ddNTPs terminate DNA synthesis.





# Improvements to Sanger sequencing and molecular methods allowed the sequencing of increasingly large genomes

- 1965 – First nucleic acid sequenced: Yeast trnA
- 1976 – First complete genome sequenced (RNA virus: bacteriophage MS2)
- 1977 – Maxam-Gilbert and Sanger DNA sequencing methods introduced and first complete DNA genome (Phage  $\Phi$ -X174)
- 1983 – PCR introduced
- 1995 – First complete cellular genome (*Haemophilus influenzae*) and eukaryotic genome (yeast) sequenced
- 2001 – Publication of the first sequenced human genomes
- 2005 – Introduction of 454 Sequencing and the NGS Revolution
- 2005 – present: Rapid evolution of NGS technology



Slide courtesy Dan Sloan.

# Next generation sequencing (NGS) ~ deep sequencing ~ high throughput sequencing (HTS)

All simultaneously sequence many molecules in parallel

## Short read sequencing

- Millions of reads
- Relatively short: ~50-300 nt (Illumina)
- Relative low error rates
- Illumina has virtually all of the market share

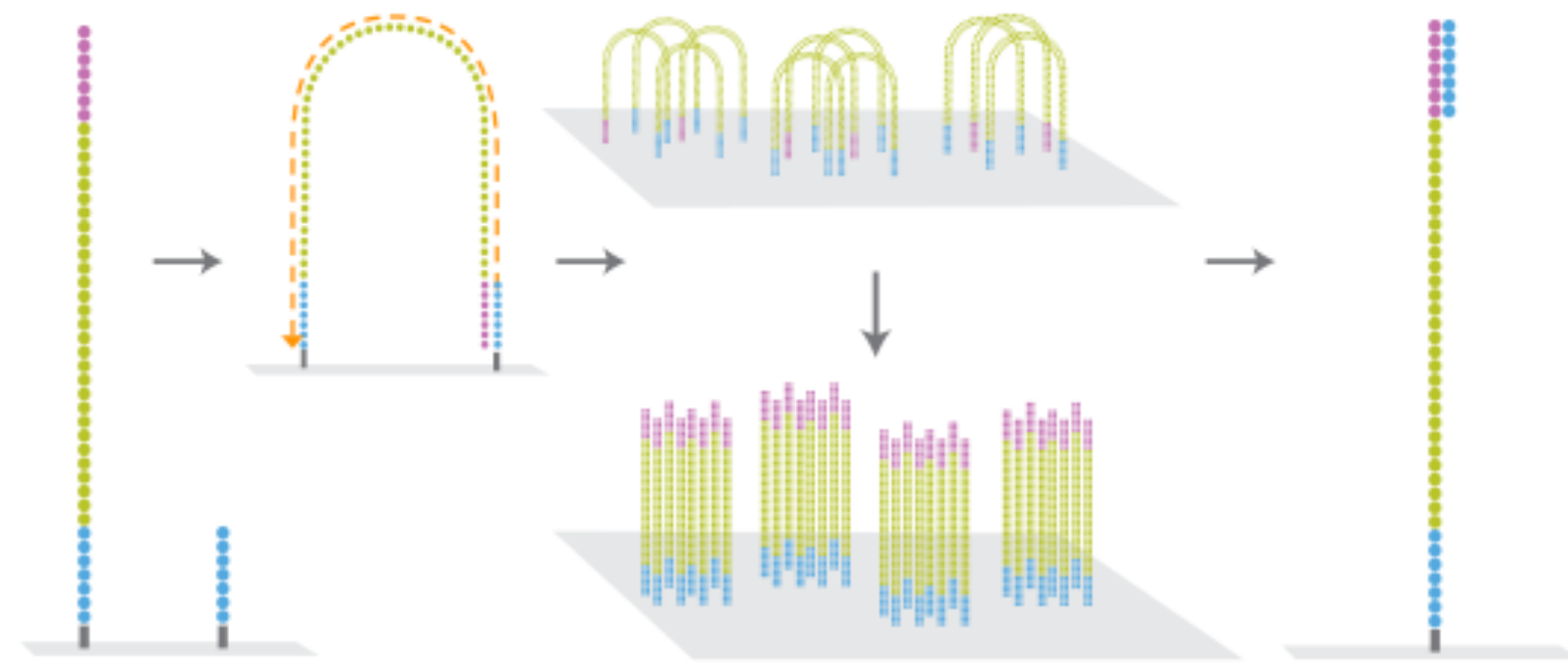
## Long read sequencing

- Fewer, longer reads
- >1 kb (PacBio), up to 100s of kb (Oxford Nanopore)
- Relative high error rates



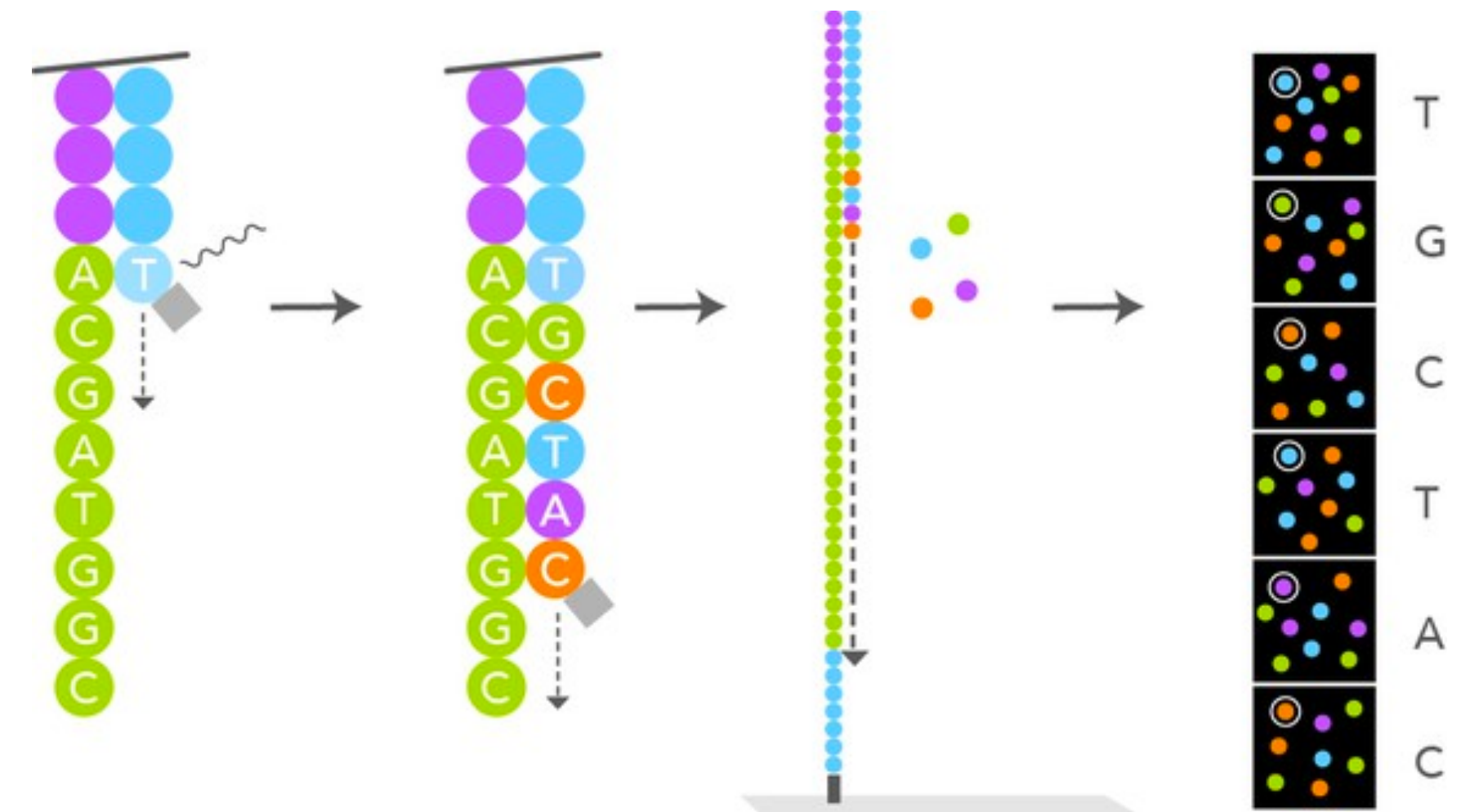


# Illumina instruments use sequencing by synthesis (SBS)



Millions of clusters per flow cell

Each cluster contains 1000s of clonal copies of a library molecule



Library molecules are sequenced by primer extension reactions that incorporate chain-terminated, fluorescent nucleotides

real raw Illumina sequencing data

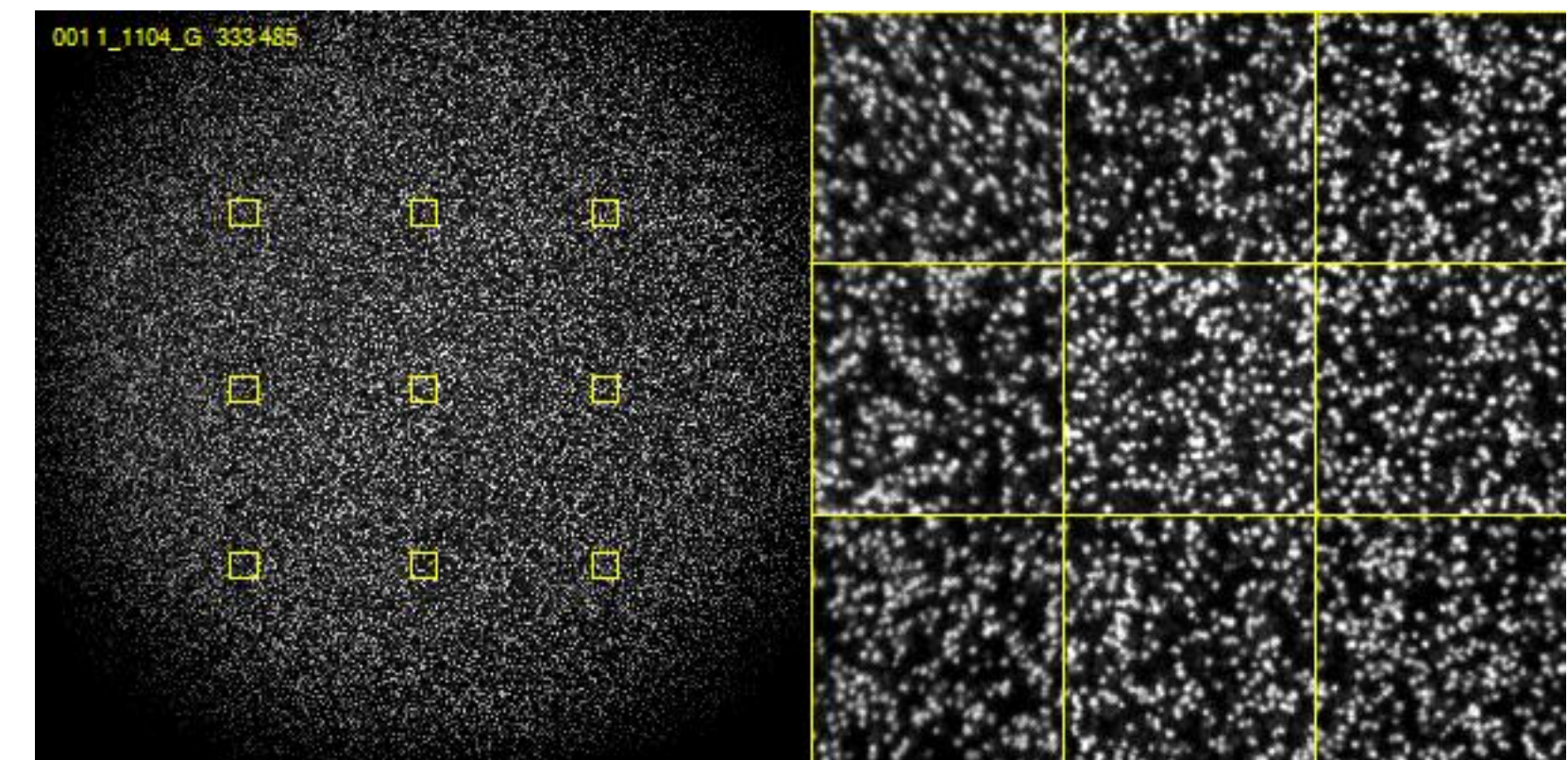
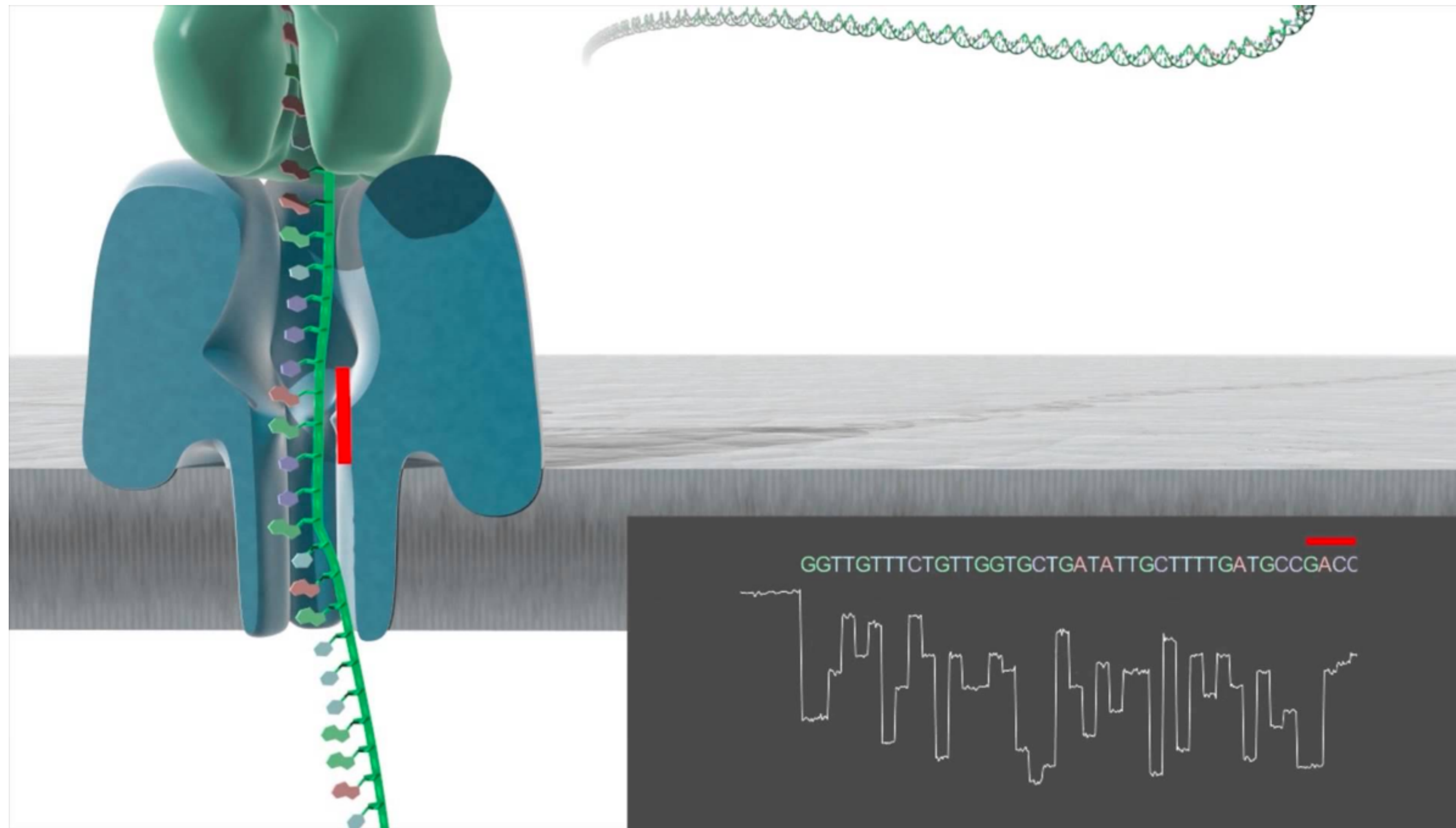


Image credit: Illumina



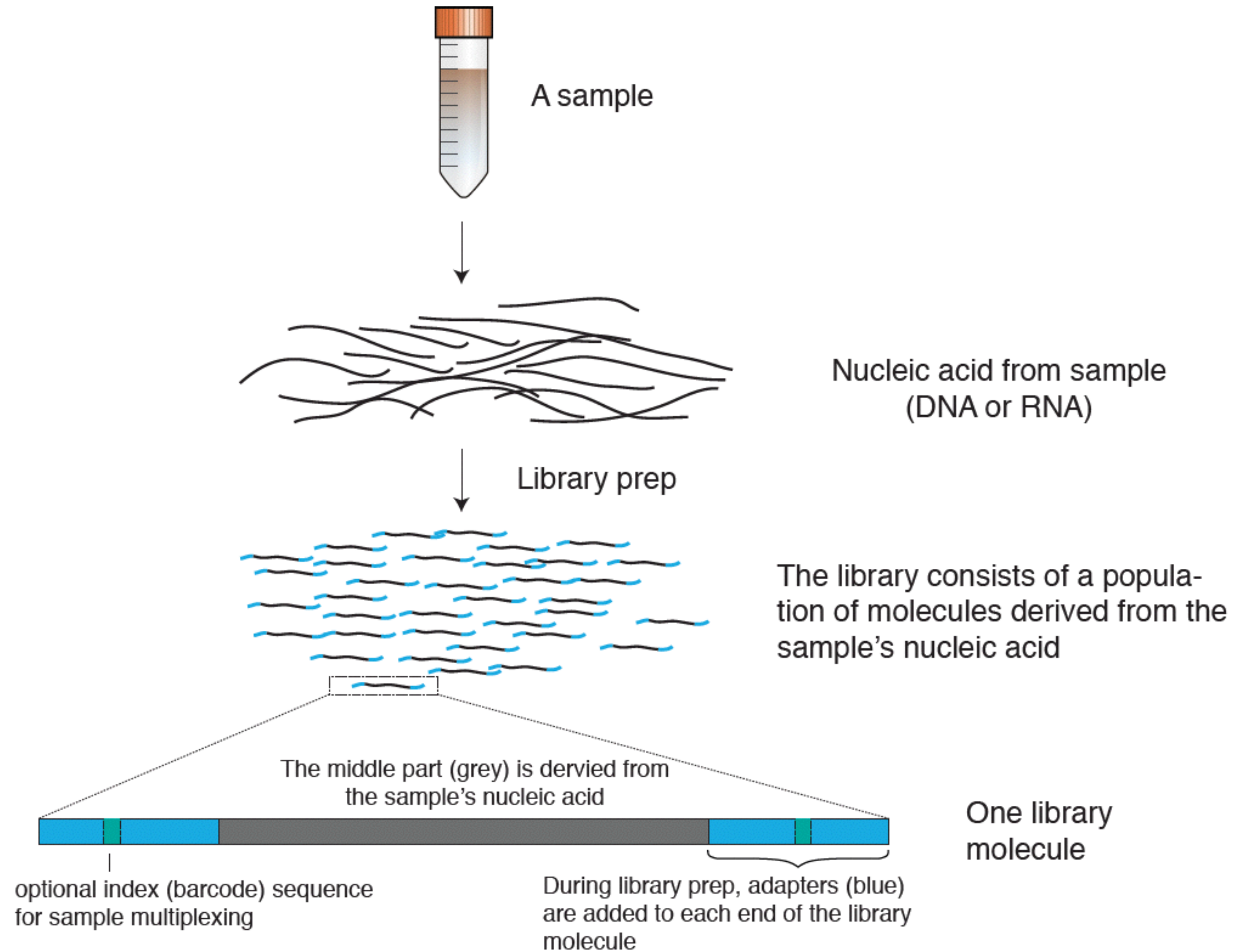
# Long read sequencers sequence single molecules



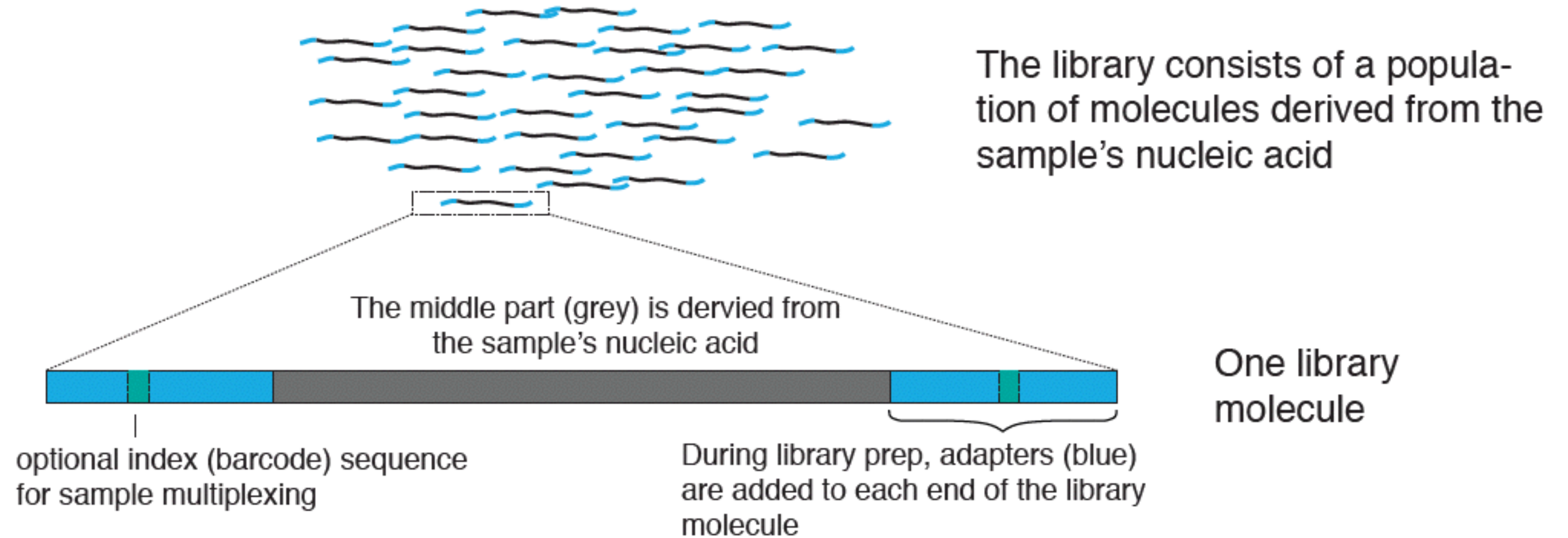
Much longer reads, but with much higher error rates



# Library prep converts nucleic acids into a form suitable to be sequenced



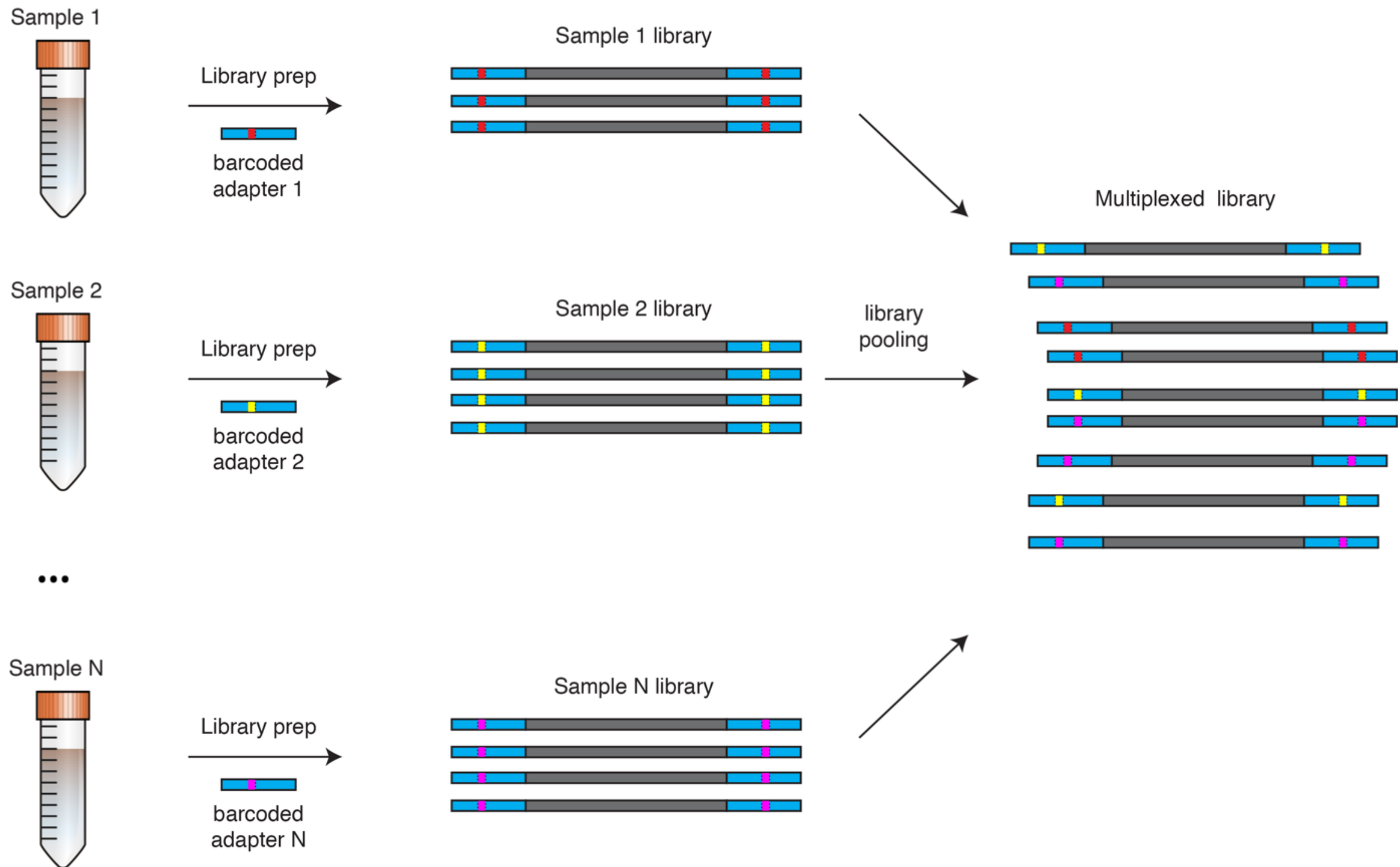
Library prep converts nucleic acids into a form suitable to be sequenced



An example Illumina library molecule

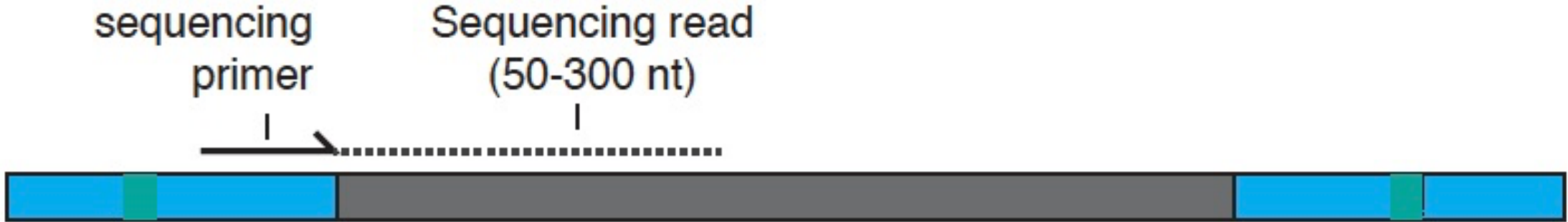


# Barcodes (or indexes) allow sample multiplexing



# Illumina sequencing produces 1-4 reads per library molecule

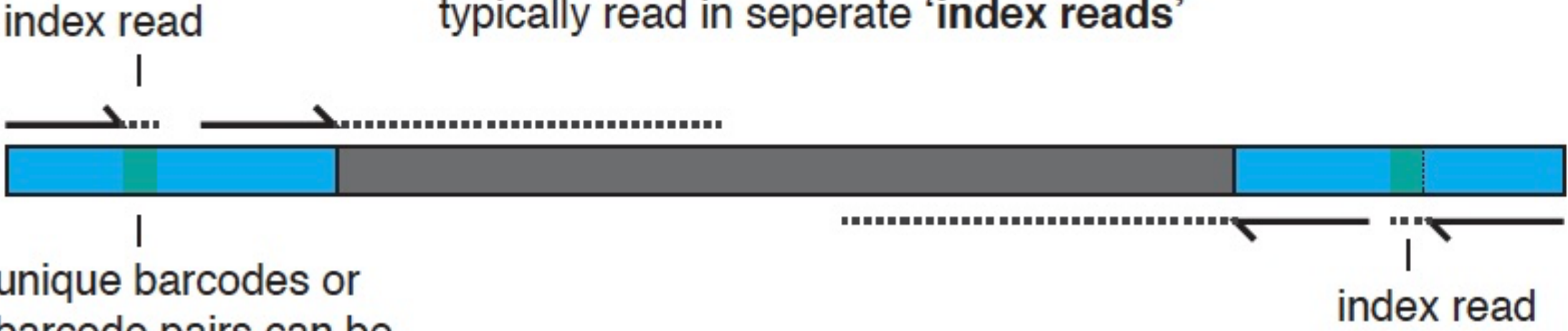
In **single end sequencing**, a library molecule is sequenced from one end



In **paired end sequencing**, a library molecule is sequenced from both end



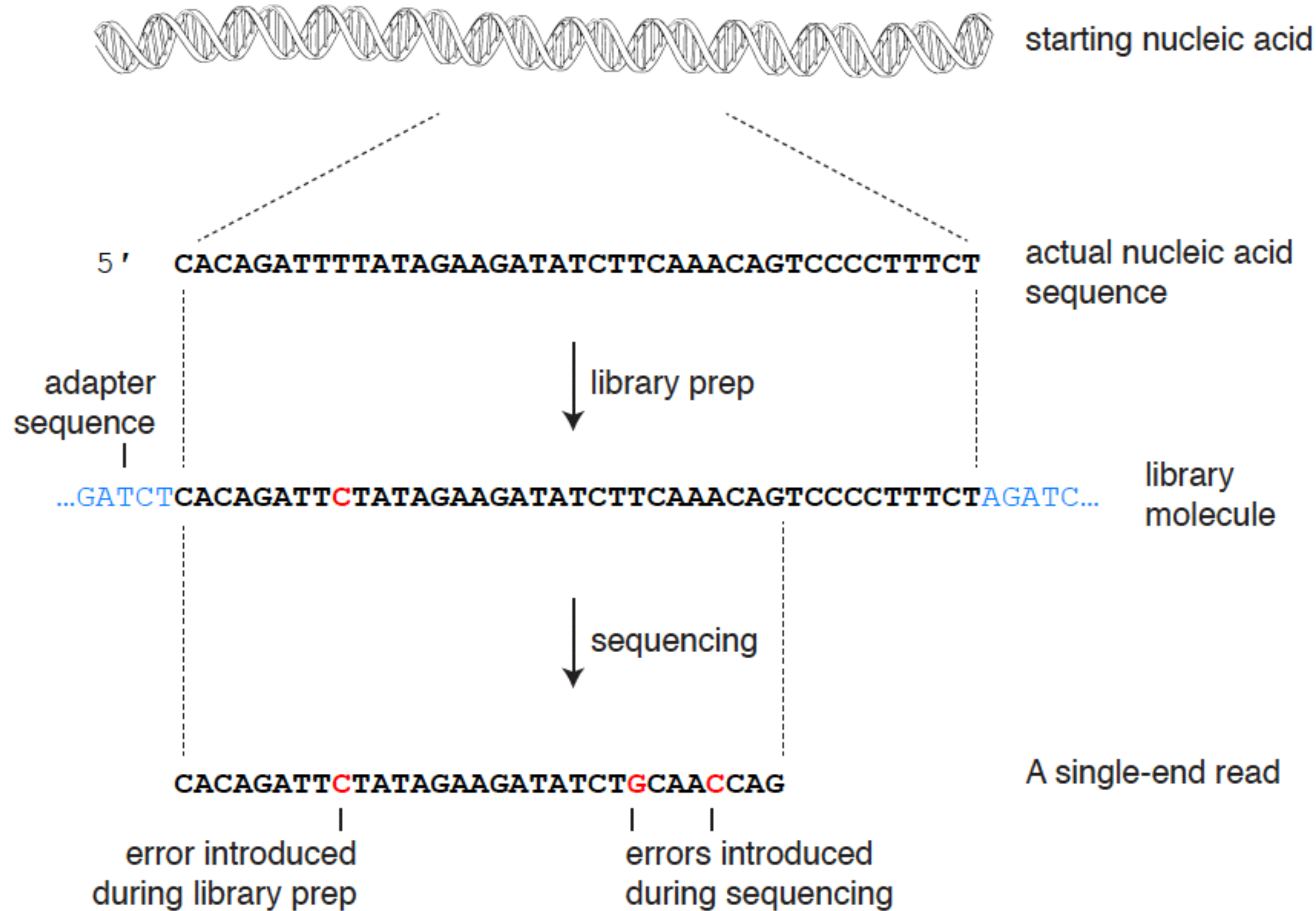
The library molecule's barcodes (indexes) are typically read in separate 'index reads'



unique barcodes or barcode pairs can be used to differentiate multiplexed samples



Reads are sub-sequences of the starting nucleic acid that often contain errors

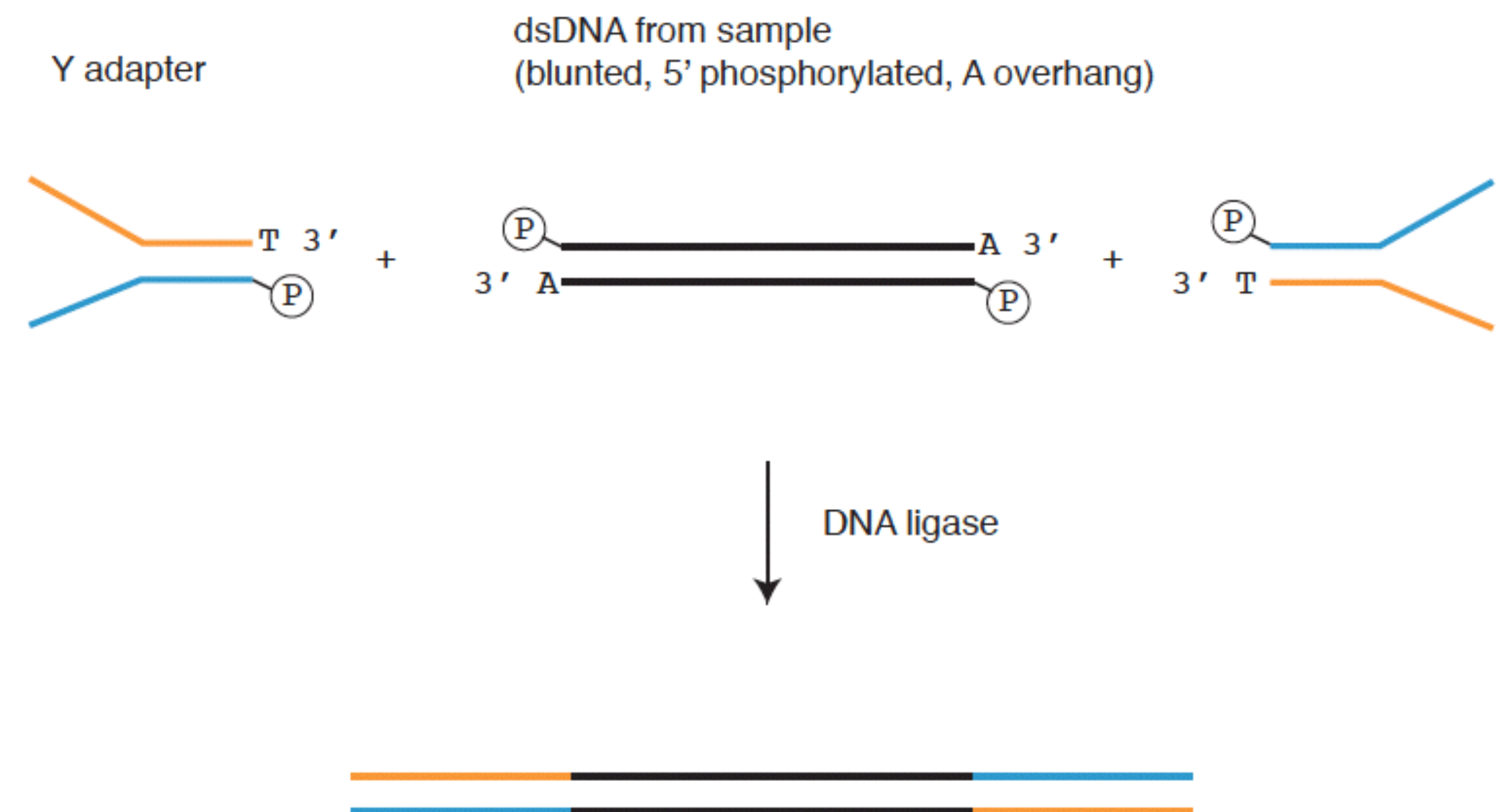


# There are many good ways to make sequencing libraries

**Common library prep steps** (not always included and not always in this order)

- Nucleic acid isolation
- Enrichment (of nucleic acid subtypes you want) or subtraction (of those you don't want)
- Nucleic acid fragmentation
- Conversion of RNA into dsDNA (for RNA sequencing)
- Addition of adapters to ends of library molecules, possibly with barcodes for multiplexing
- Library amplification
- Pooling of multiplexed samples
- Library QC / quantification

Adapters can be added to sample-derived dsDNA by ligation

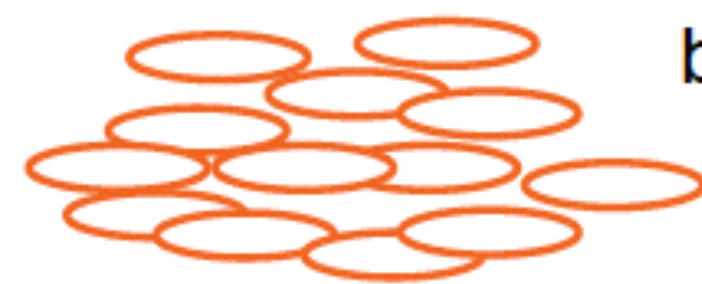
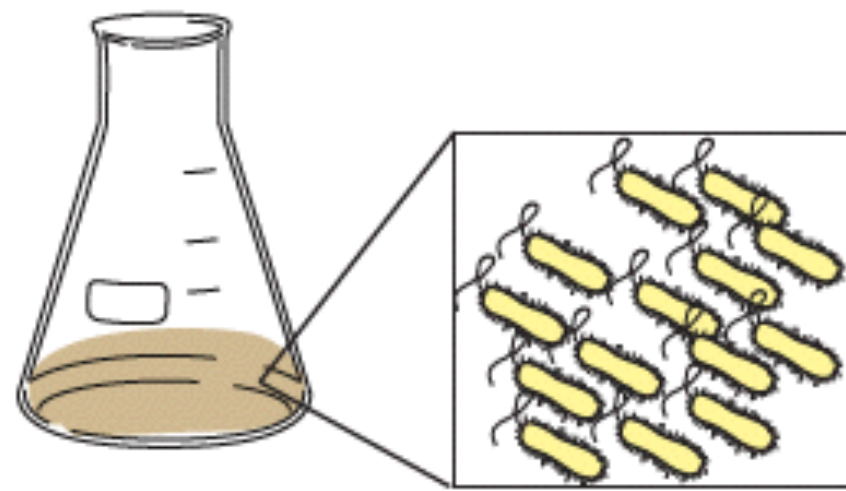




How you make a library determines what type of sequencing you're doing

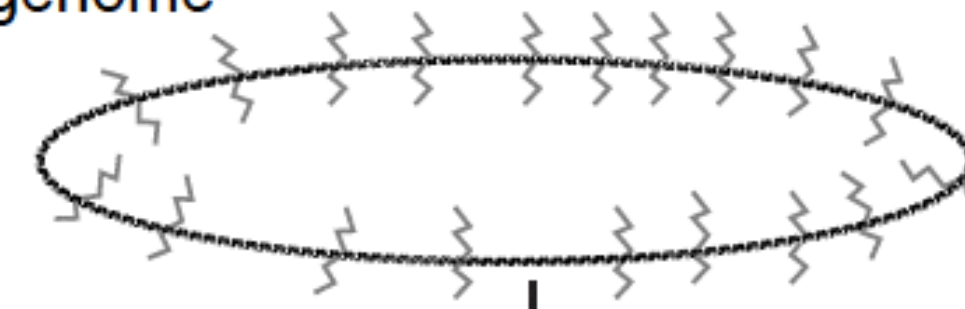
For instance, if you make a 'shotgun library' from a single organism, you're doing **whole genome sequencing** (WGS)

bacterial isolate



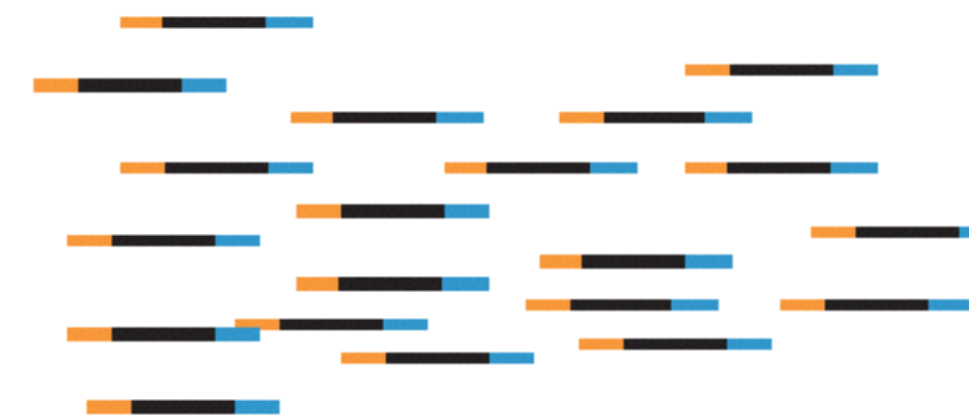
bacterial genome

bacterial genome



shotgun sequencing samples the entire genome

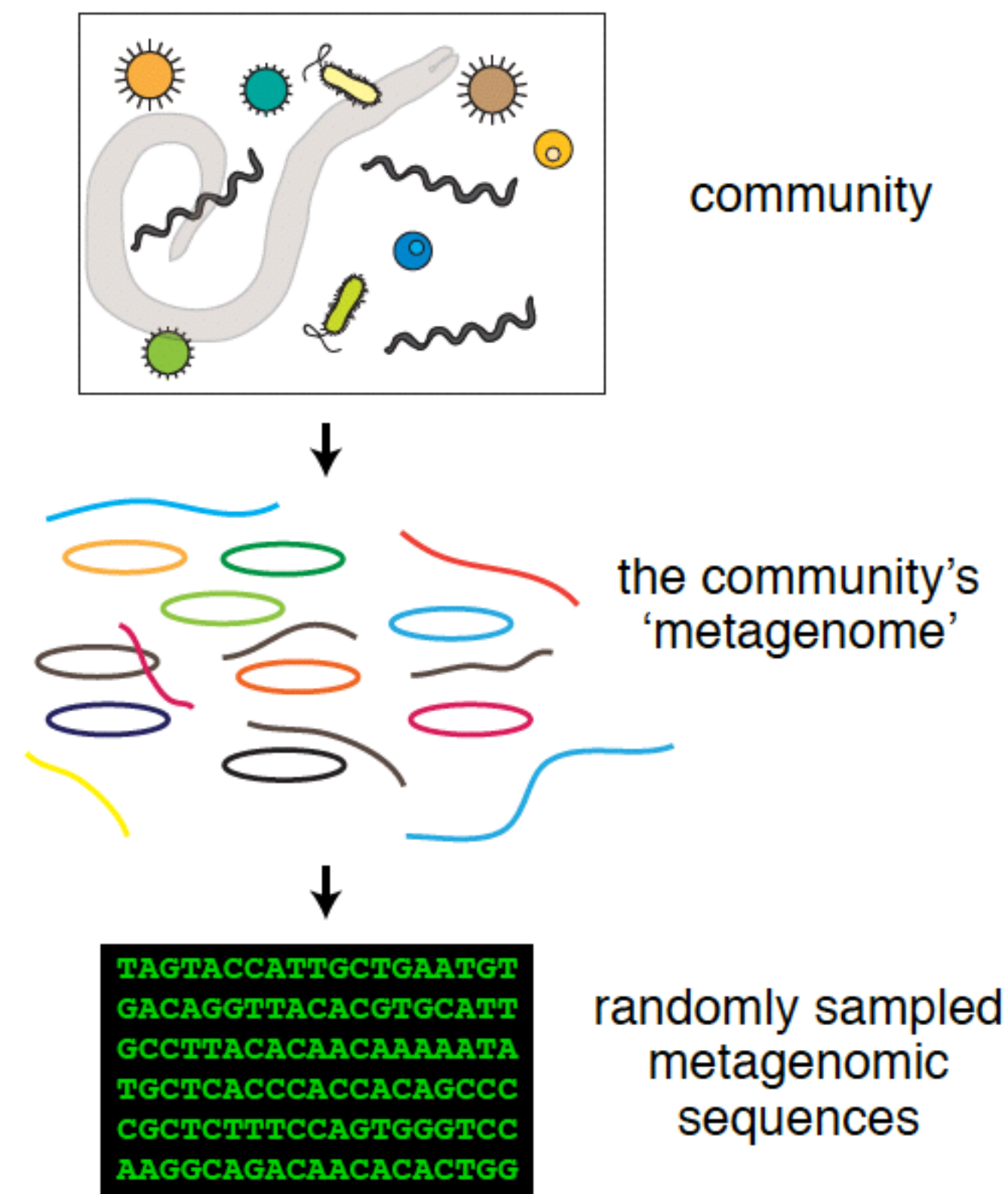
random fragmentation and adapter addition



shotgun library molecules contain random bits of the genome

# How you make a library determines what type of sequencing you're doing

For instance, if you make a 'shotgun library' from a community DNA sample, you're doing **metagenomics**

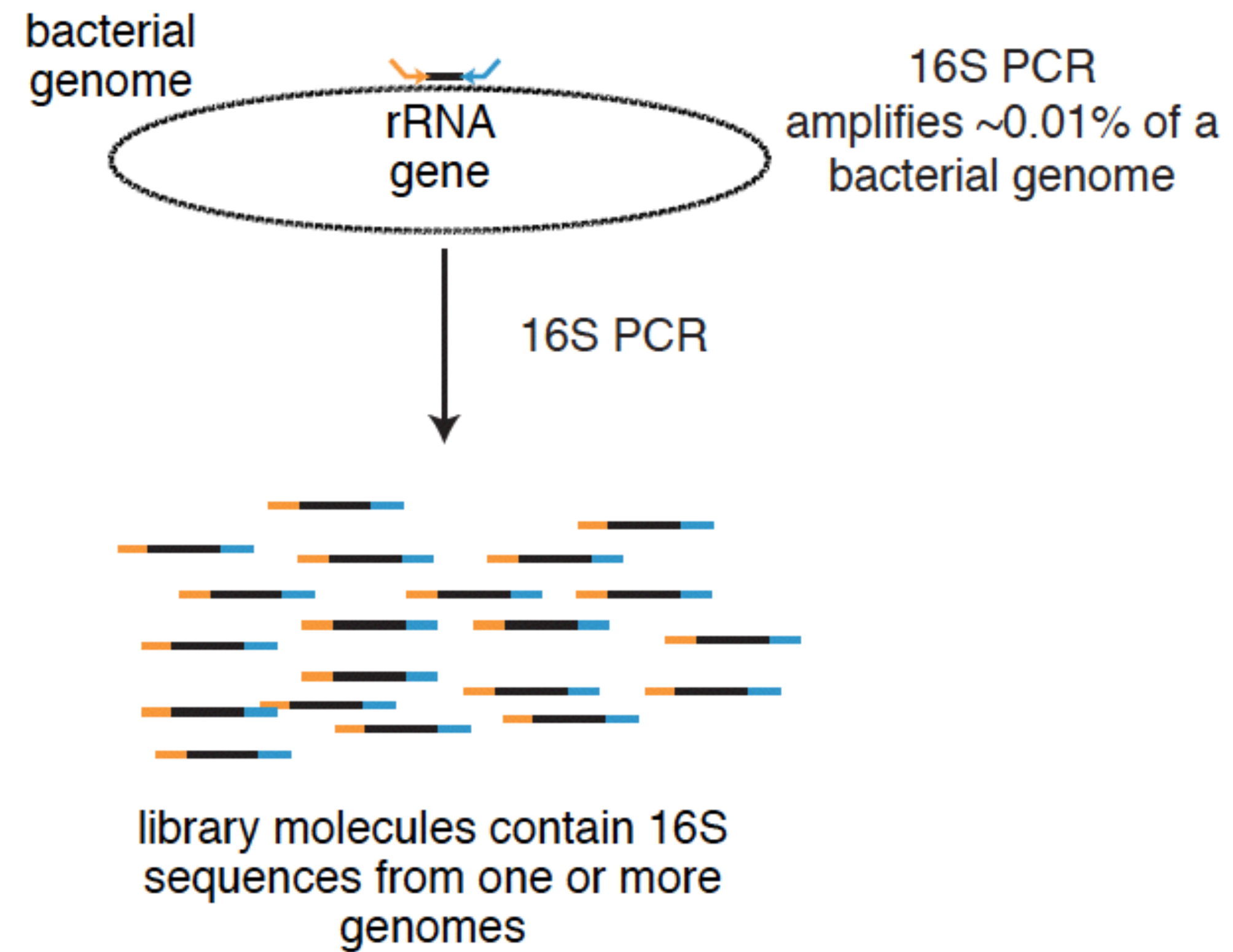


The library preparation methods are identical to those used for whole genome sequencing. You're just starting with a different kind of DNA sample.



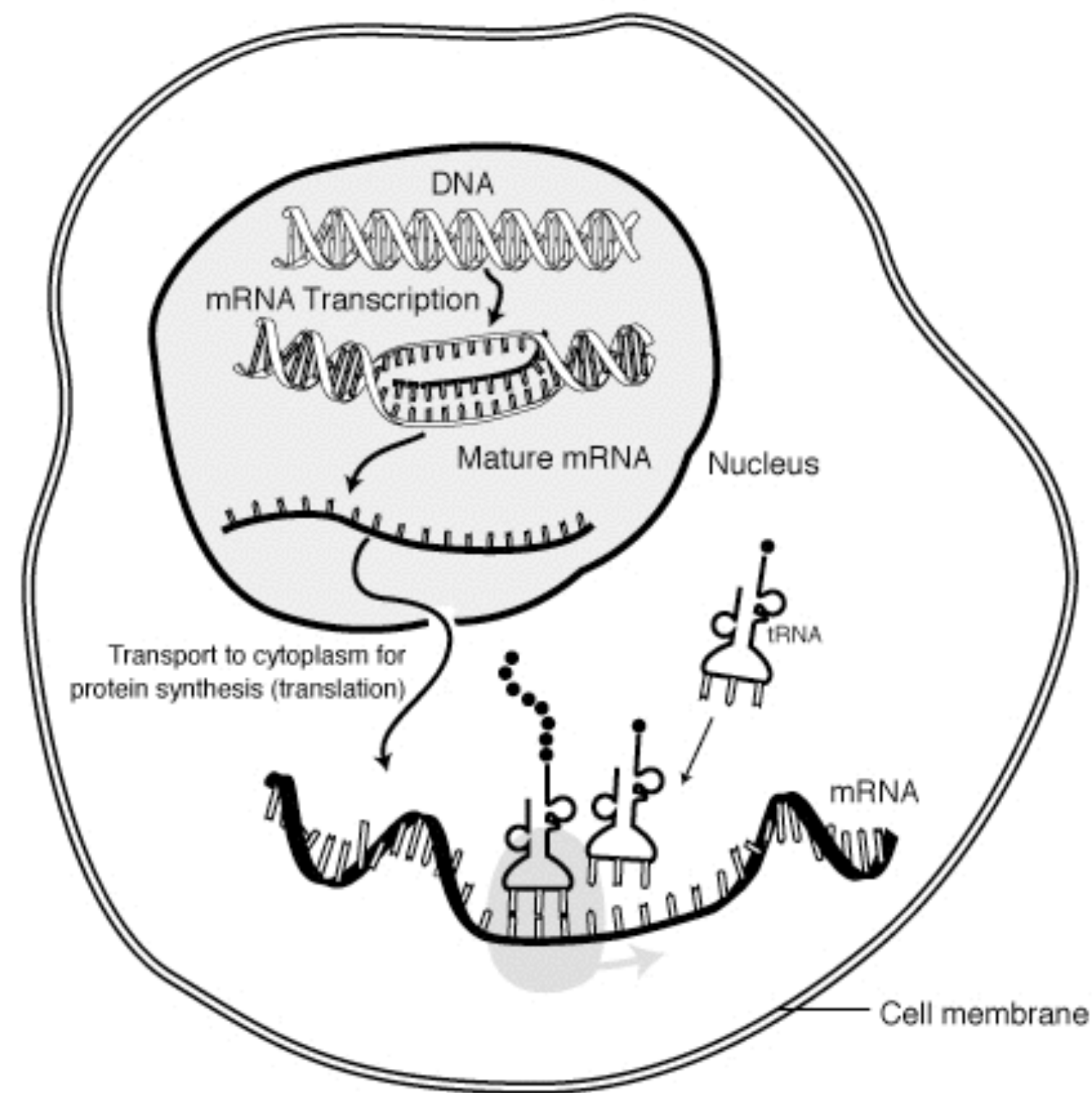
# Microbiome sequencing often means **16S rRNA** sequencing

16S sequencing is one type of  
'amplicon sequencing'



How you make a library determines what type of sequencing you're doing

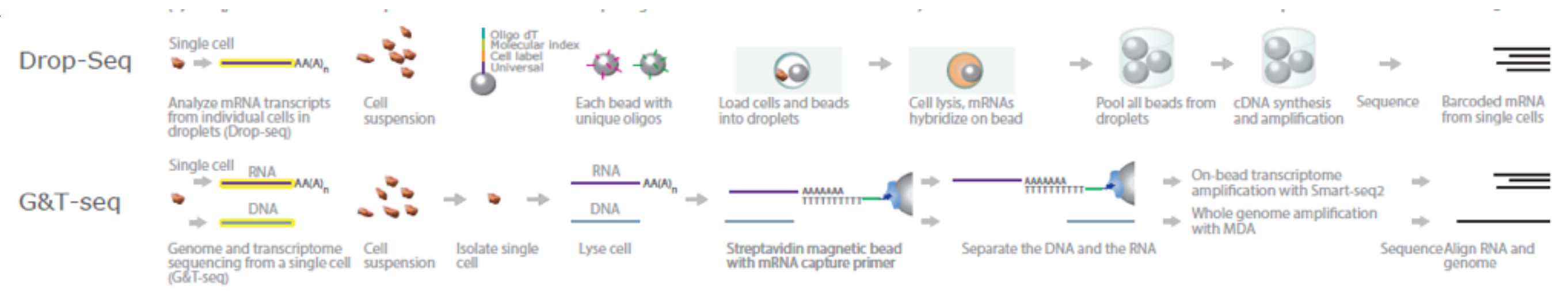
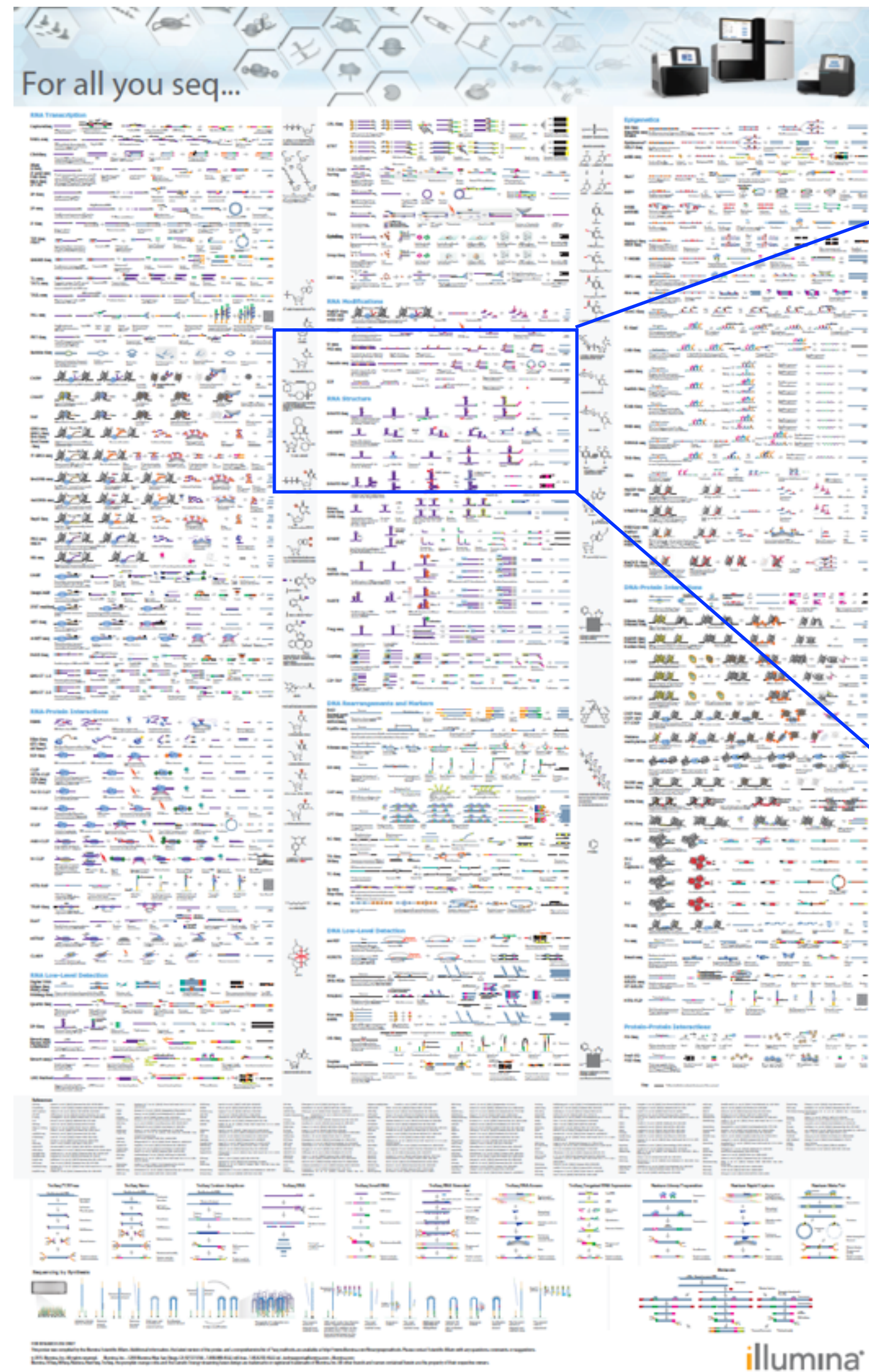
If you make a library from mRNA, that is **RNA-Seq** (transcriptome sequencing)



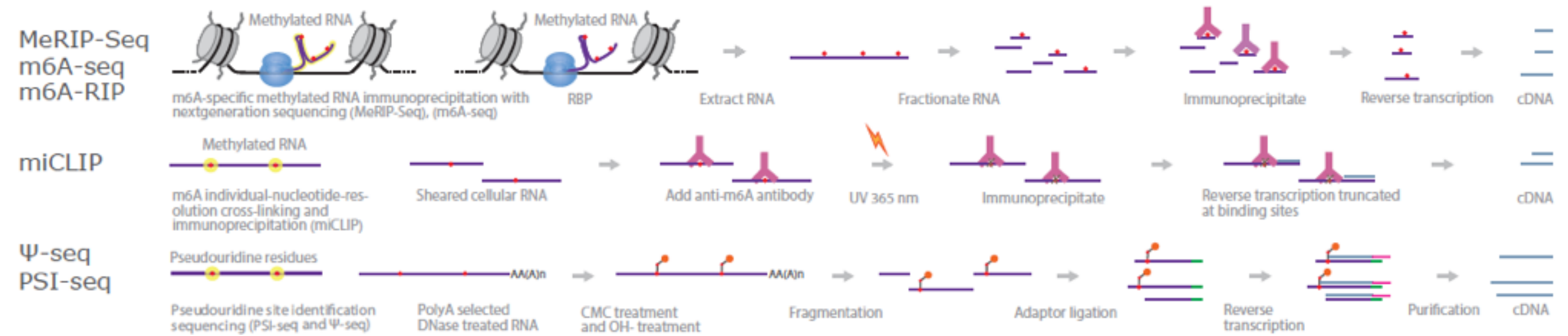
The abundance of reads from a particular mRNA is proportional to that mRNA's abundance in the cell



There are **5 billion** ways to make libraries and to do sequencing (all have names that end in -seq)



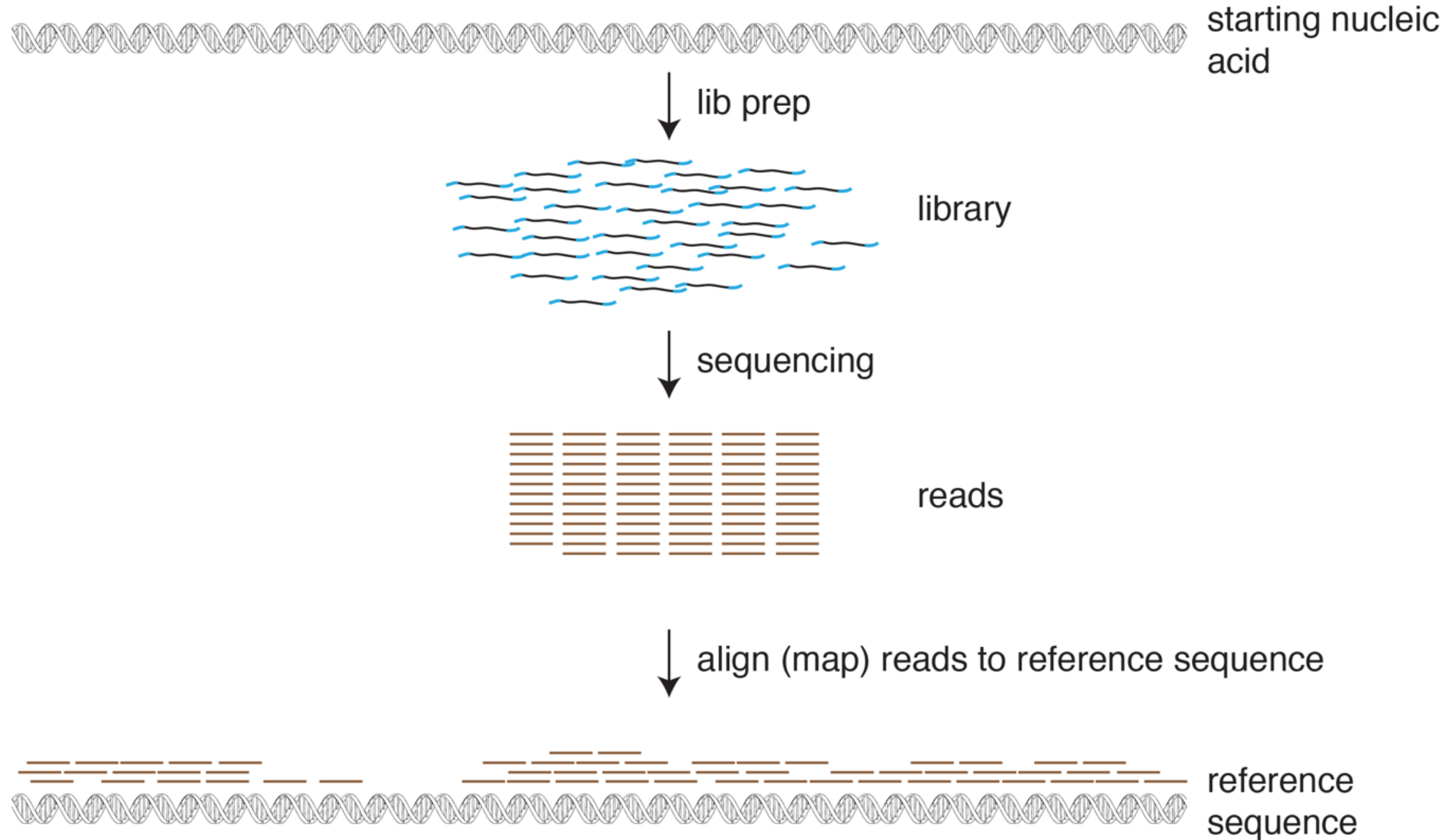
### RNA Modifications



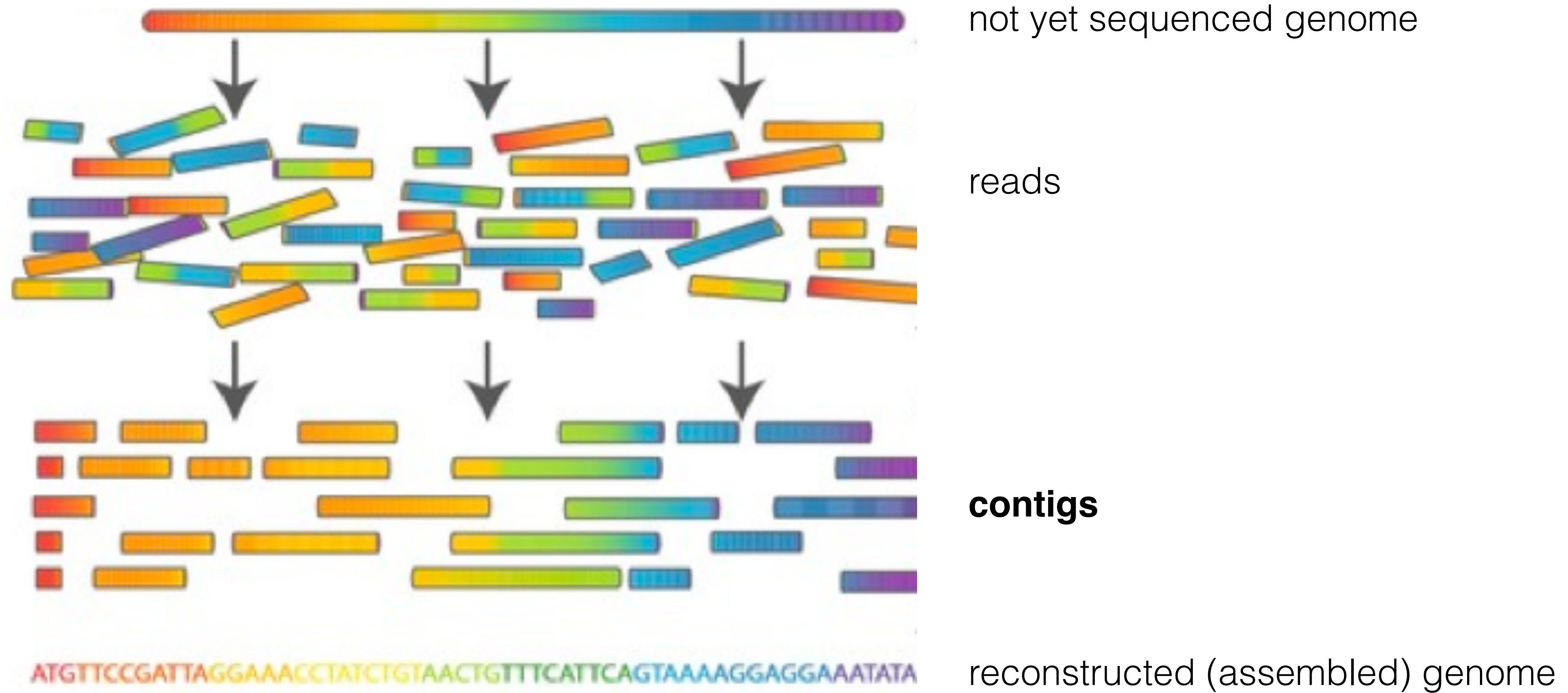
They're all variations on a few themes. Don't let it overwhelm you. Most sequencing is of a few simple types, and it's better to focus on the Biology and experimental design.



**Mapping** is the process by which sequencing reads are aligned to the region of a genome from which they derive.

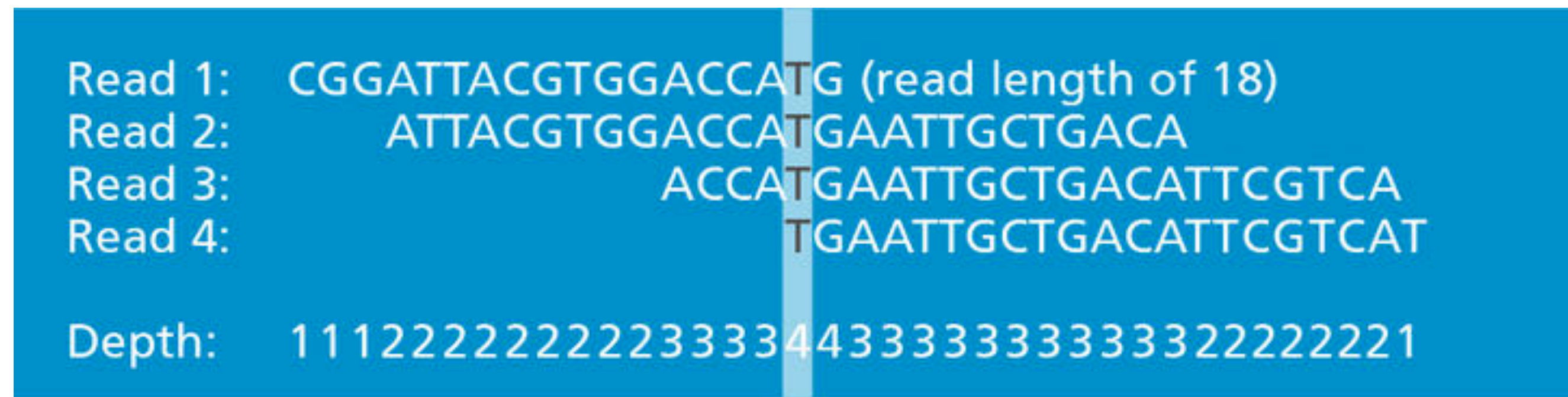


(De novo) **assembly** is the process of trying to reconstruct a genome sequence from reads



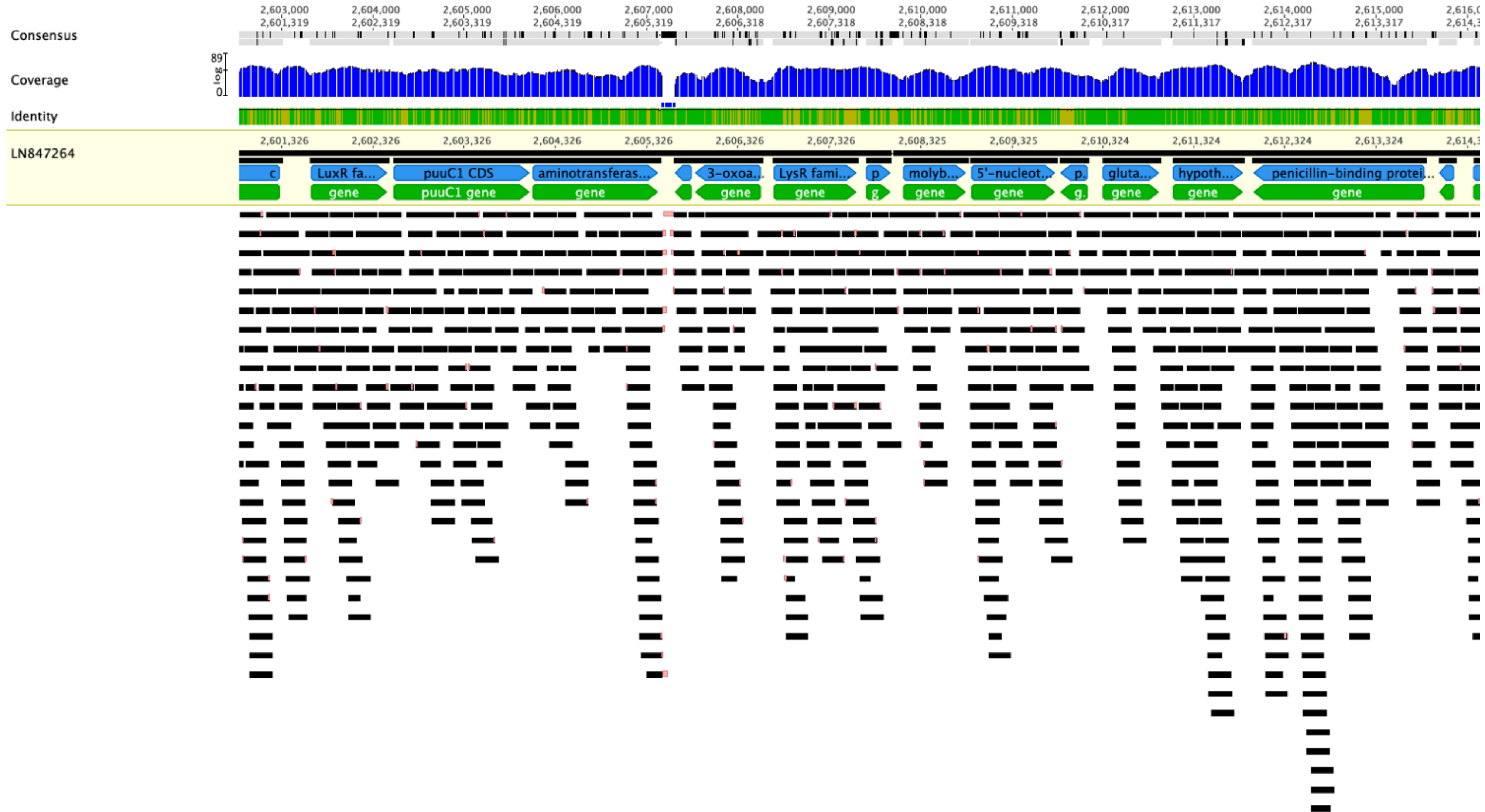


**Coverage** is the number of individual reads that support a particular nucleotide in an assembled (reconstructed) sequence or that align to a particular nucleotide in a reference sequence



coverage is often referred to as 'depth' or 'depth of coverage'

# Coverage from WGS of a bacterial isolate



Questions?

Is there genomics or sequencing jargon about which you're not certain?