

# Outline

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- Introduction to RNA-seq
- Sample preparation
- Quality control
- Read alignment
- Differential gene expression
- Data visualization and plotting

# Regulation of gene expression

## Regulation of transcription:

- Transcription factors
- Histone modifications
- DNA methylation

## Regulation of RNA processing:

- Polyadenylation
- Splicing
- Capping
- RNA export

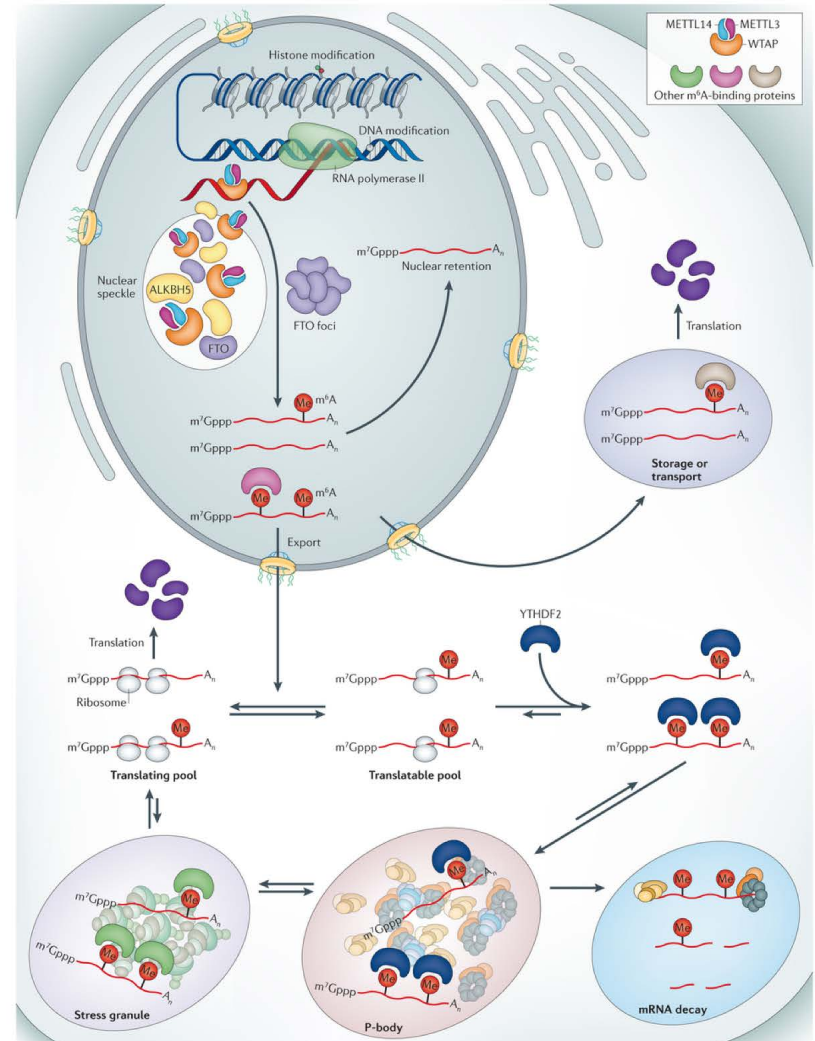
## Regulation of translation:

- mRNA decay
- Translational repression
- Sequestration

## Posttranslational regulation:

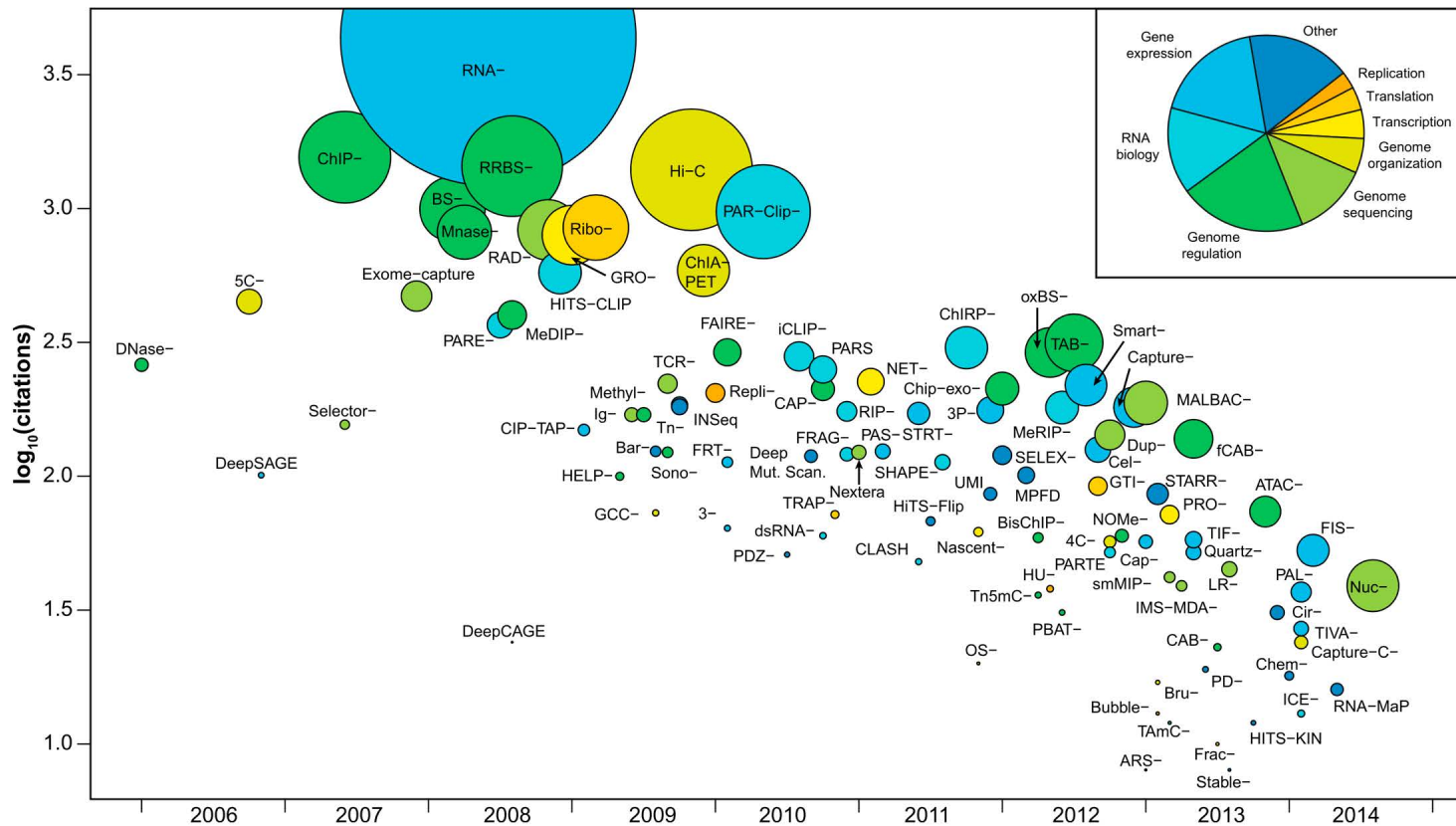
- Chemical modifications (e.g. phosphorylation)
- Protein turnover (proteolysis)

RNA-seq measures steady state mRNA levels and RNA sequence composition





# RNA-seq is the most common HTS application



Reuter et al 2015

# Sample preparation

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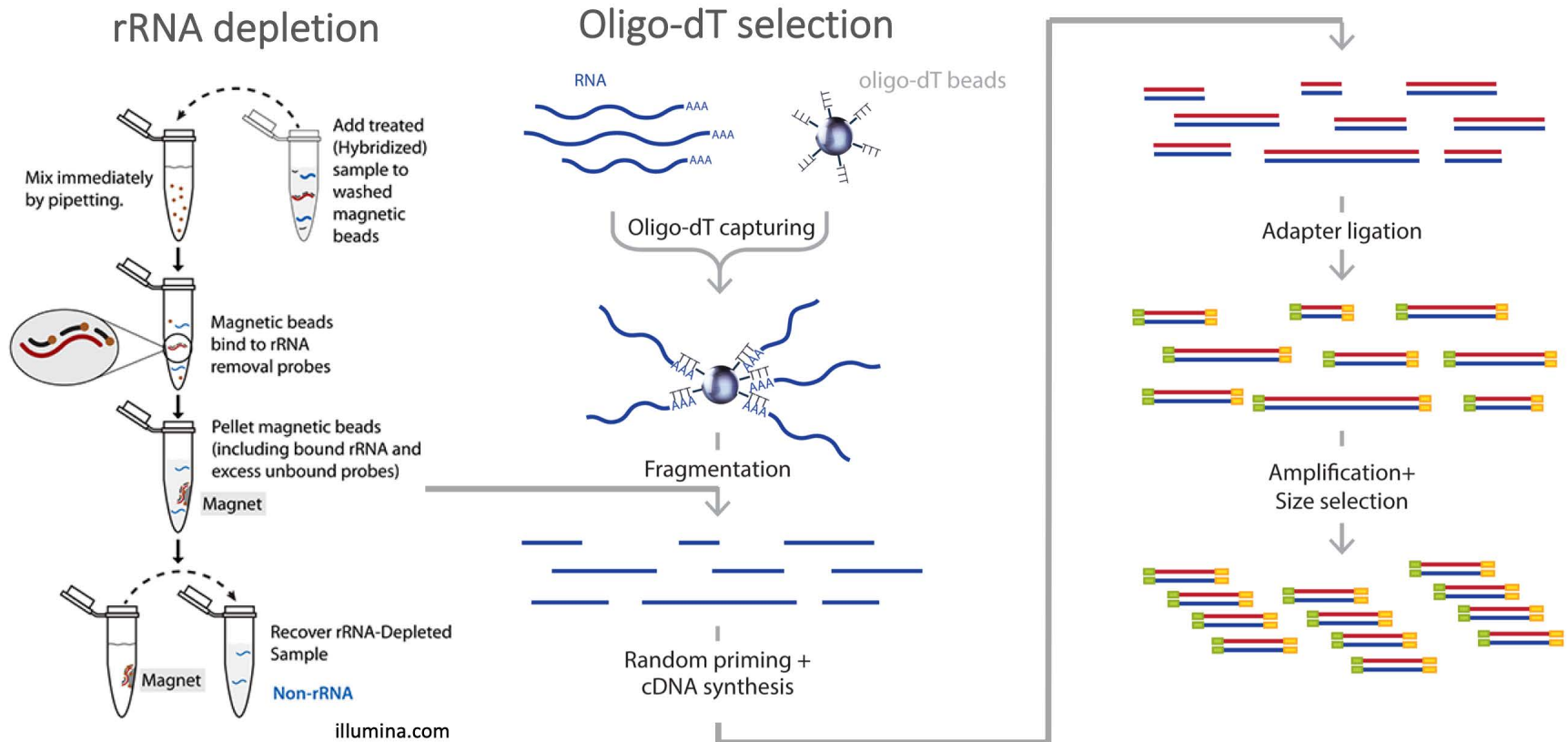
- Use high-quality RNA as starting material.
- Minor differences between samples can have a substantial impact on gene expression.
- 3-4 biological replicates is the default but not ideal for every situation.
- Some recommended kits for standard RNA-seq:
  - NEBNext Ultra II Directional RNA Library Prep Ki
  - Illumina kits

# Sample preparation

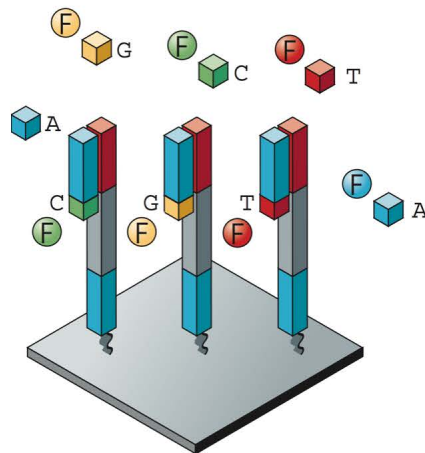
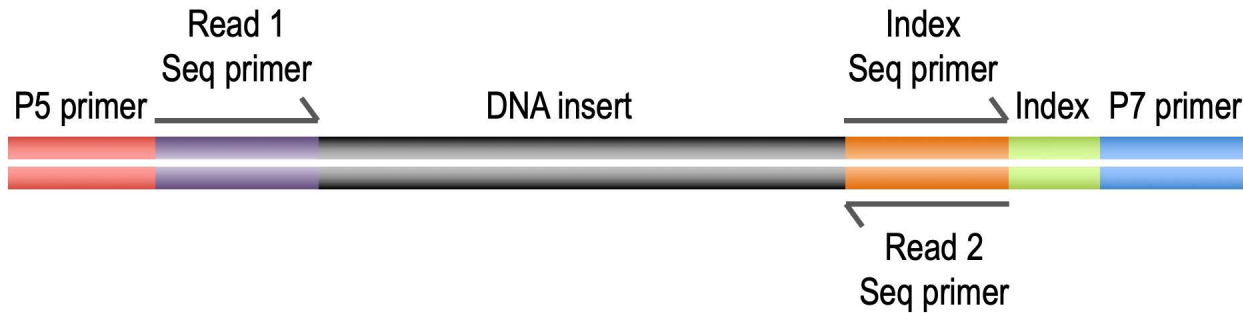
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- Starting RNA
  - Typically 1-5 ug of high-quality total RNA is ideal.
- Sequencing depth
  - Typically you want 20-30 million high quality reads/library.
- Considerations
  - Strand specific (default is yes)
  - Single-end or paired-end (single-end is typically sufficient)
  - Long reads vs short reads (short Illumina reads, 50-150 nt, are usually sufficient)
  - rRNA depletion or oligo-dT
  - Low quantity/single cell

# RNA-seq library preparation



# Library composition



Metzker, M.L. (2010) NRG



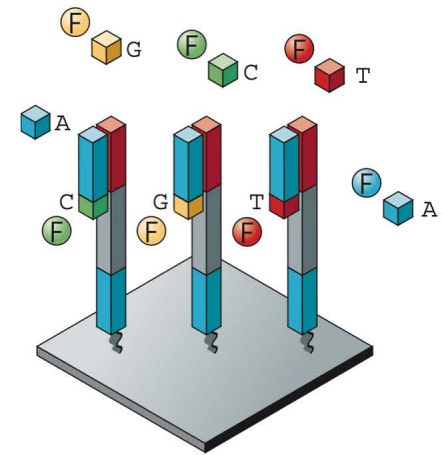
HiSeq 2500

# FASTQ format

## Index sequence

```

Read 1 [ 1 @D64TDFP1:248:C50DMACXX:5:1101:1241:2095 1:N:0:ATCACG
        2 CACCGCCCGTCGCTATCCGGGACTGGAATTCTCGGGTGCCAAGGAACTCCA
        3 +
        4 CCCFFFFFFHHHHHJJIJGHJJJJJJJJJJGGGFFFEABDHHHFHFF@@DD>
Read 2 [ 1 @D64TDFP1:248:C50DMACXX:5:1101:1371:2154 1:N:0:ATCACG
        2 TCAATATTTGCATAGGGTATCTGGAATTCTCGGGTGCCAAGGAACTCCAGT
        3 +
        4 CCCFFFFFFHHHHHJJJJGFHJJJJJJJJJJJJJJFHHIJJJHGHJFGHJJI
Read 3 [ 1 @D64TDFP1:248:C50DMACXX:5:1101:1461:2205 1:N:0:ATCACG
        2 GAAAGACGTCTTCTAGATTATGGAATTCTCGGGTGCCAAGGAACTCCAGT
        3 +
        4 CCCFFFFFFHHHHHJJJJJJJJJJJJJJJJJJJJJJHJJJJGIIJFGIJJJ
    
```



Metzker, M.L. (2010) NRG

Line 1: sequence ID, description, and index; begins with @

Line 2: sequence; contains only A, C, T, G, and N

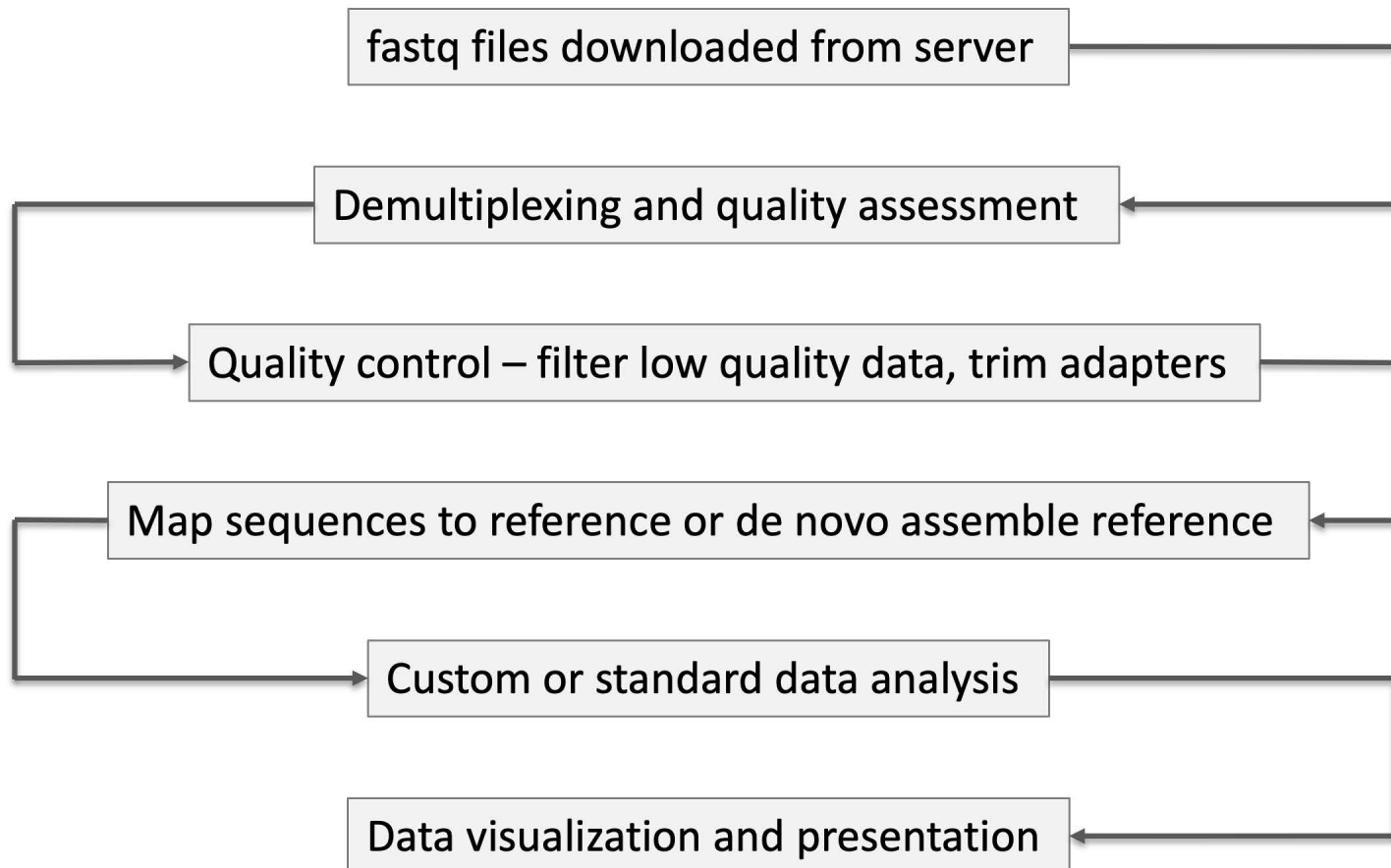
Line 3: optional sequence ID; begins with +

Line 4: signal quality of each base, cryptic code, phred 33 or 64



# Data analysis workflow

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# Quality control

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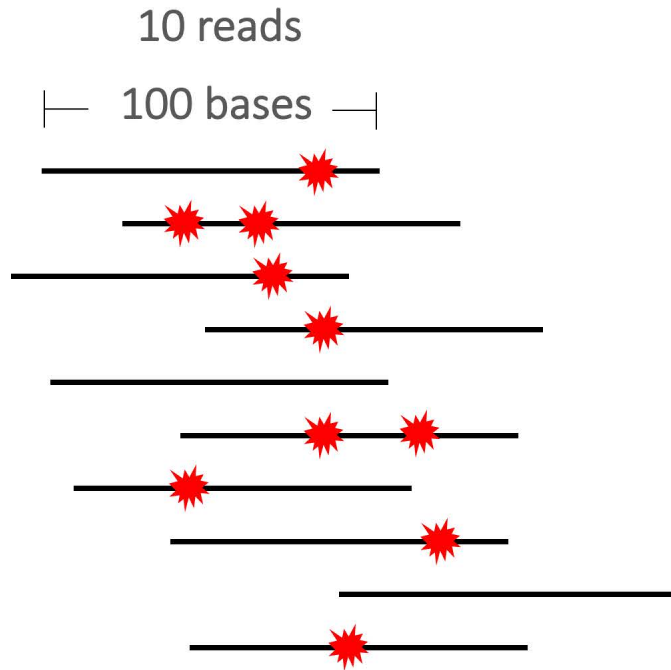
## Assessing Read Quality

Phred quality score: a measure of the quality of base calling:

$Q = -10 \log(P)$  where  $P$  is the error probability

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10,000	99.99%
50	1 in 100,000	99.999%
60	1 in 1,000,000	99.9999%

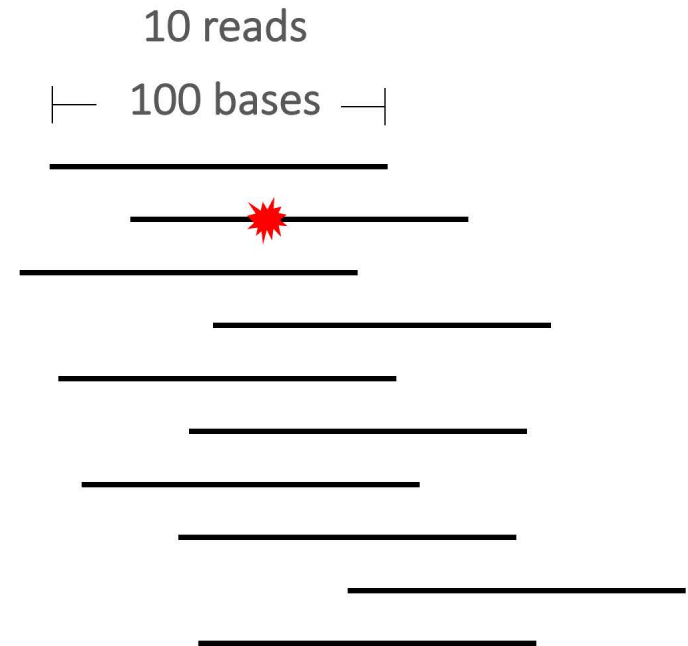
# Quality control



$$P = 0.01$$

$$Q = 20 \text{ (Q20)}$$

$$Q = -10 \log(P)$$



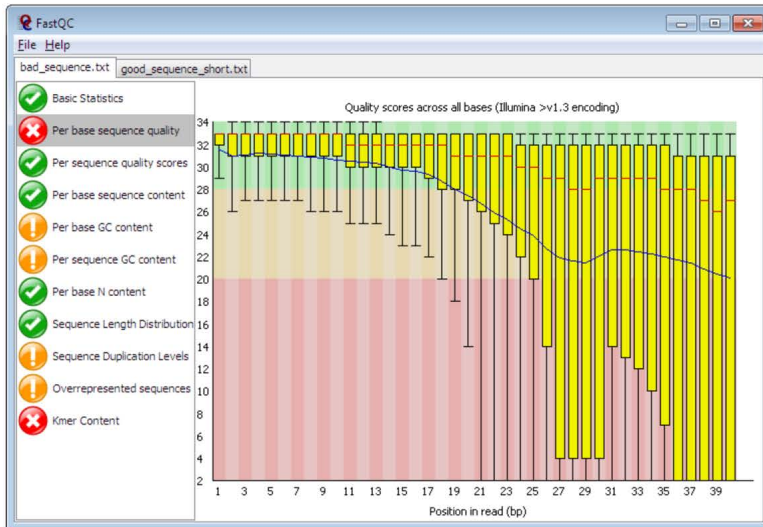
$$P = ?$$

$$Q = ?$$

Q30 is a common quality threshold or quality criterion

# Quality control

FastQC: a GUI tool for assessing the quality of high-throughput sequencing data.



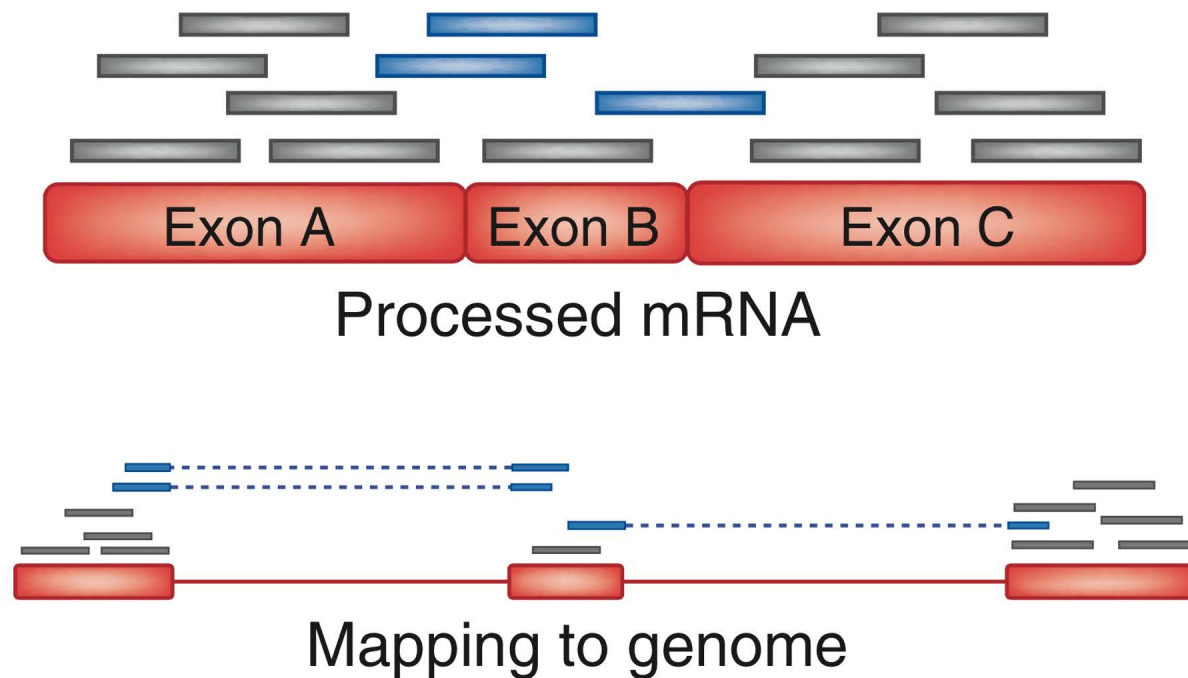
Trimmomatic: software for trimming adapter sequences and low-quality bases from sequencing reads.

THE USADEL LAB



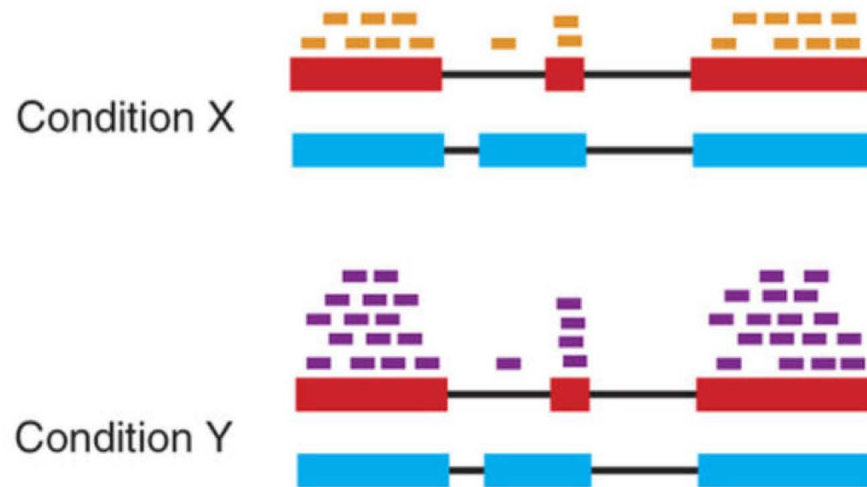
# Aligning reads to mRNAs

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Trapnell et al (2009)

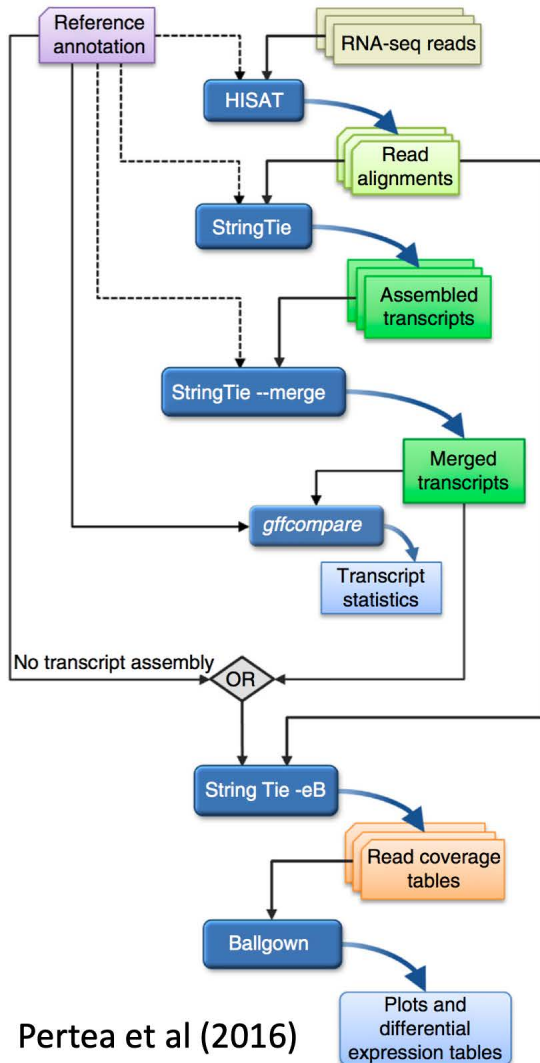
# Differential gene expression



Trapnell et al (2010)

How many reads align to each gene in condition X vs condition Y?

# RNA-seq pipelines



No reference genome? Use Trinity to assemble transcripts

Other mRNA aligners: Star, GNSAP, Tophat2

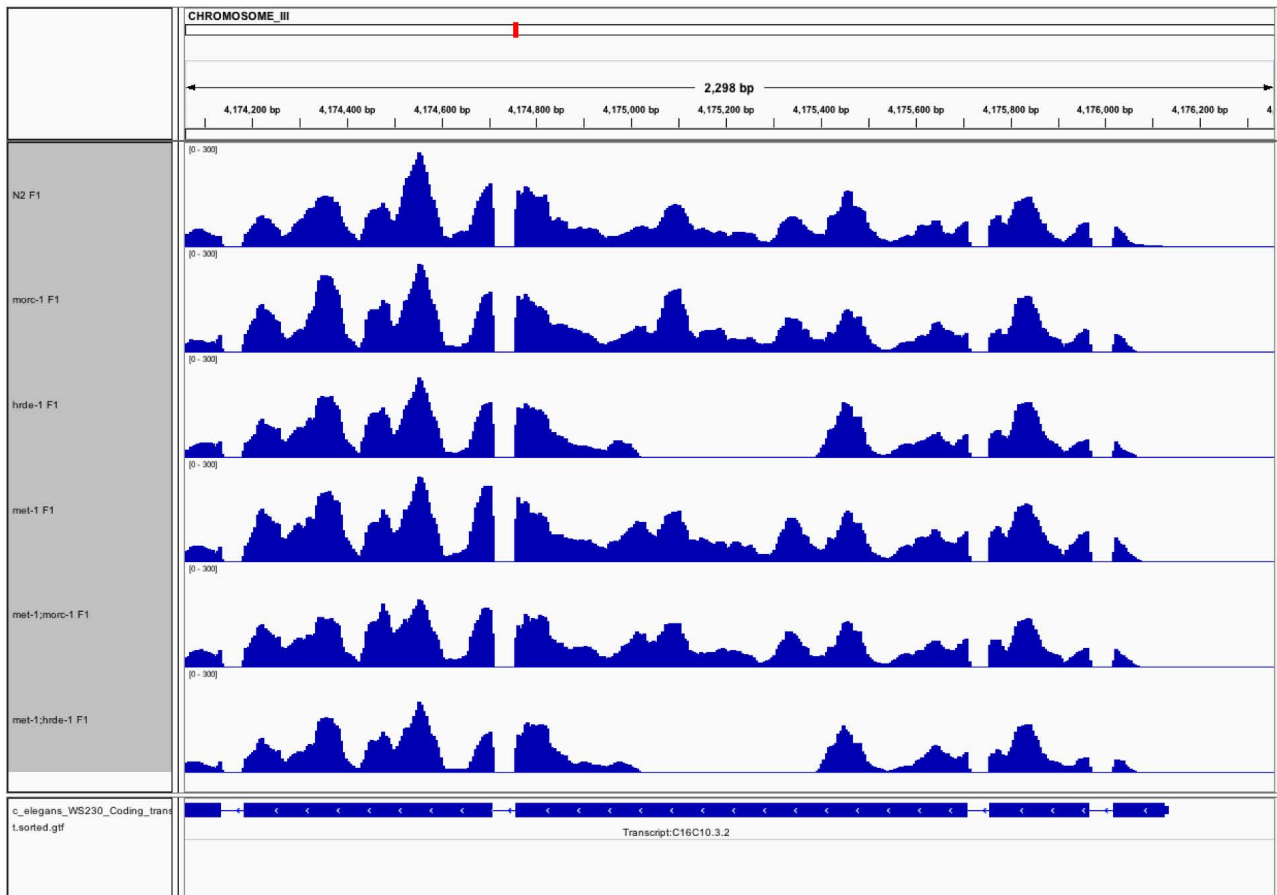
Other abundance estimators: RSEM, htseq-count, kallisto (alignment free estimates), salmon (alignment free estimates)

Other common DE software: DESeq2, edgeR, cuffdiff

Various GUIs and R-based tools for drawing plots

# Genome browsers

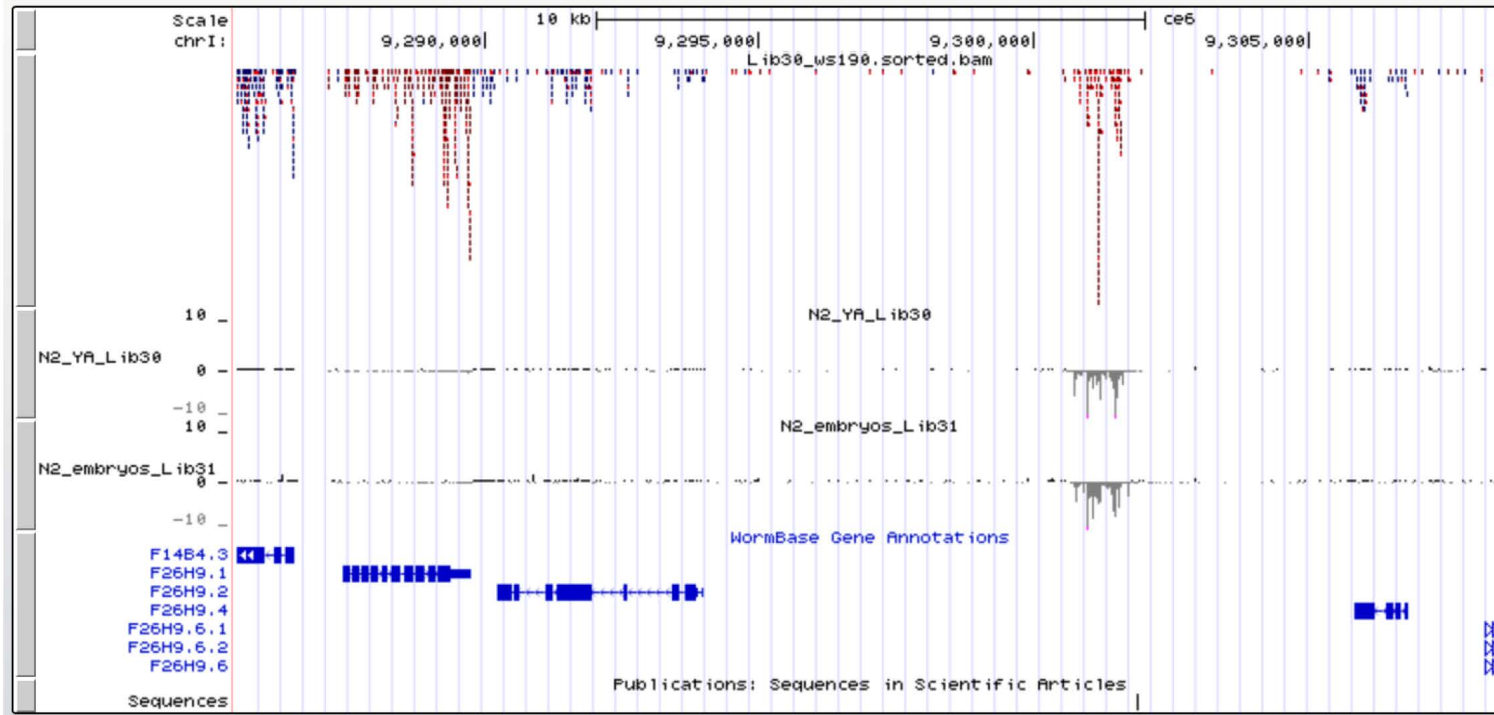
## Integrative Genomics Viewer (IGV)





# Genome browsers

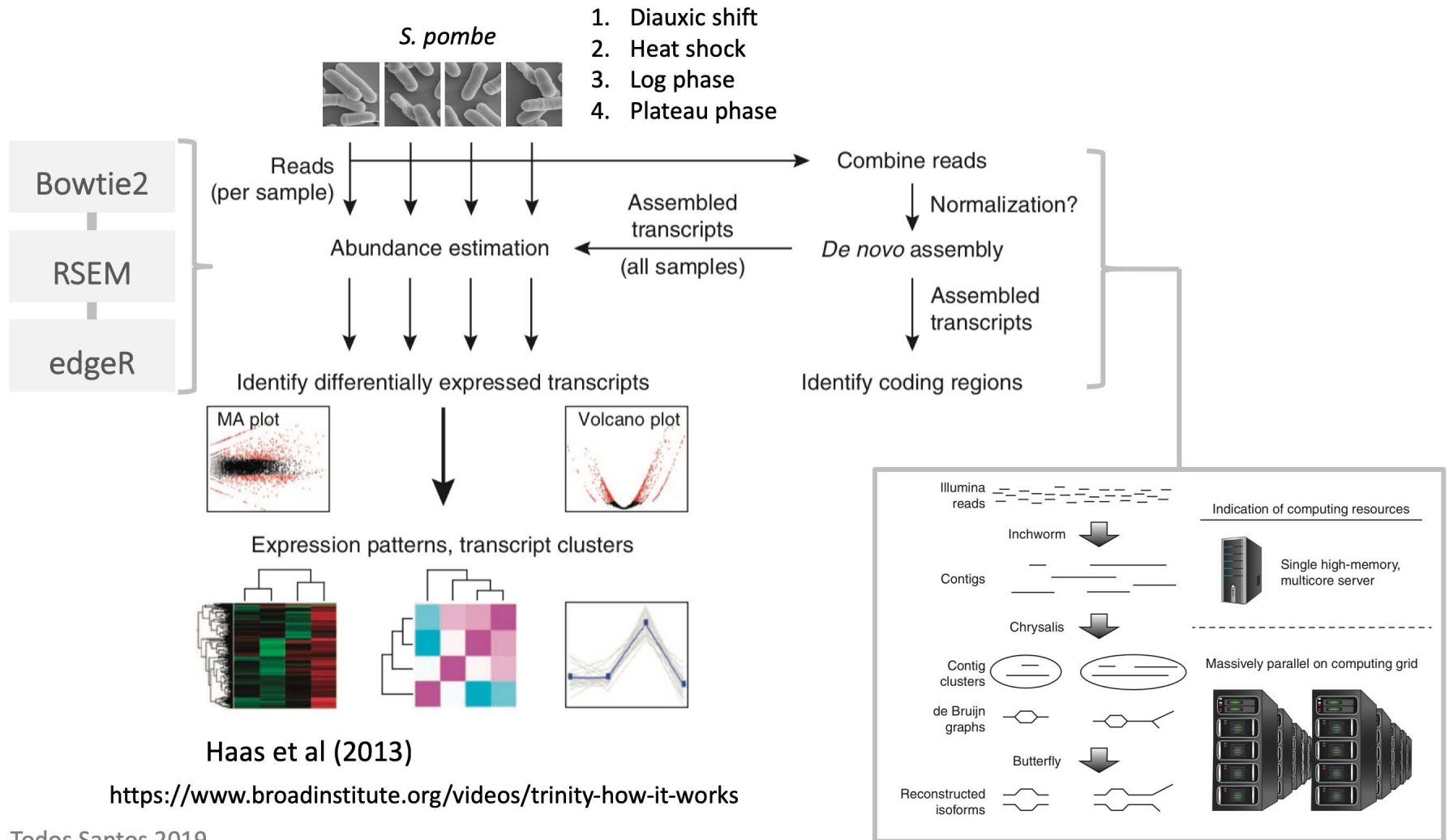
## UCSC Genome Browser



<https://genome.ucsc.edu>



# Trinity workflow



# Functional annotation

## Trinotate



Pfam



eggNOG  
version 3.0



RNA-Seq → Trinity → Transcripts/Proteins → Functional Data → Discovery

Automated Higher Order Biological Analysis

<http://trinotate.github.io>