Practical Choices in Sequencing Projects



Frequently Asked Questions in NGS Library Construction

- **Platform**. Which Illumina sequencing platform is best? Or should I be using long-read technologies?
- **Read-Lengths**. How many sequencing cycles should I run?
- **Paired-End.** Should I do paired-end or single-read sequencing?
- **Read Number.** How many reads should I generate?
- **PCR**. How many PCR cycles should I do and which polymerase should I use?

Short-Read vs Long-Read Next-Generation Sequencing Techniques

Short-Read Sequencing Long-Read (Single Molecule) Sequencing



Oxford Nanopore

PacBio

Under what circumstances would you want to use short-read vs. long-read sequencing?

Illumina Sequencing Platforms

	Clusters	Max Read-	Max	Cast	Bacterial	Eukaryotic
	(millions)	Length	Output (Gb)	Cost	Genomes	Transcriptomes
MiniSeq	25	150 bp	7.5	\$1,500	15	1.5
MiSeq	25	300 bp	15	\$1,530	30	3
NextSeq 500 (mid)	130	150 bp	40	\$1,650	80	8
NextSeq 500 (high)	400	150 bp	120	\$4,240	240	24
HiSeq 4000 Lane	300	150 bp	90	\$1,925	180	18
NovaSeq S4 Lane	2500	150 bp	750	\$9,100	1500	150



Principles when Choosing Read Lengths

- Read lengths are defined by the cycle number on an Illumina run (1 bp per cycle).
- Advantages of longer reads
 - Cost per bp declines
 - MiSeq V2 50 cycles -- \$747 (\$996/Gb)
 - MiSeq V2 300 cycles -- \$958 (\$213/Gb)
 - MiSeq V2 500 cycles -- \$1066 (\$142/Gb)
 - Better for distinguishing among repetitive sequences in assembly/mapping
- Disadvantages
 - Worthless if your inserts are short
 - Additional sequence is not "independent" (e.g., for quantifying gene expression)
 - Basecall quality diminishes with read length.

Read Quality

Quality declines with increasing cycle number because amplicons within clusters get out of phase.



Single-Read vs. Paired-End



Advantages of paired-end runs

- Cost per bp declines
 - NextSeq V2 1x150 cycles -- \$2650 (\$44/Gb)
 - NextSeq V2 2x150 cycles -- \$4240 (\$35/Gb)
- Better for distinguishing among repetitive sequences in assembly/mapping

Disadvantages

- Worthless/redundant if your inserts are short
- Additional sequence is not "independent" (e.g., for quantifying gene expression)

How Many Reads Do I Need?

de novo genome assembly

- 100x sequence coverage (e.g., 5 Mb genome → 500 Mb total sequence data)
- Longest paired-end reads available
- But consider PacBio/Nanopore

Genome re-sequencing (SNP and indel variant calling)

- 20x and 35x for haploid and diploid genomes, respectively
- · Longest paired-end reads available
- Low error rate technologies

RNA-seq for measuring gene expression (with ref genome)

- 36 million reads to get reliable quantification for 80% of human genes with FPKM > 10. (ENCODE 2011 PLoS Biol. e1001046)
- Short single-end reads

Table 2 | Representative read counts for location-based approaches

Techniques	Read counts in representative studies	Refs
DNasel-seq and FAIRE-seq	20–50 million	79
CLIP-seq	7.5 million; 36 million	89,90
iCLIP and PAR–CLIP	8 million; 14 million	105, 106
CHiRP and CHART	26 million	72
4C	1–2 million	92
ChIA–PET	20 million	107
5C	25 million	108
Hi-C	>100 million	94
MeDIP-seq	60 million	109
CAP-seq	>20 million	110
ChIP-seq	>10 million per sample (point source); >20 million per sample (broad source)	79

Sims et al. 2014 (Nature Reviews Genet. 15: 121-132)

How Many Reads Do I Need?

https://genohub.com/next-generation-sequencing-guide/

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	Designing Your Next Generation	Sequencing Run			
Overview >	Back to NGS Handbook While we've standardized the way sequencing runs are ordered across	s all platforms, customization is			
Type of Run	required when determining the run parameters to achieve your experimental goals. If you're new to high throughput sequencing and have questions about how you should design your sequencing run, fill out our free consultation form and we'll get in touch with you to help.				
Read Length					
Number of Reads	We highly recommend that use Genohub's NGS Matching Engine as a great tool to determine the right amount of sequencing capacity on various instruments and easily explore different options. Simply enter				
Depth of Coverage (DNA)	your specifications and instantly see services that match your output r				
Depth of Coverage (RNA)	Type of Run – Single Read (SR) or Paired End (PF)				
Replication, Randomization and Multiplexing	With single read runs the sequencing instrument reads from one end of a fragment to the other end. Paired end runs read from one end to the other end, and then start another round of reading from the				
Poor Quality Sequencing Run	opposite end. Single read runs are faster, cheaper and are typically su studies such as RNA-Seq or ChIP-Seq.				
Library Preparation	Paired end runs give additional positioning information in the genome, making it a good choice for de novo genome assembly as well as making it easier to resolve structural re-arrangements such as deletions, insertions and inversions. Experiments designed to study splice variants, epigenetic modifications				
Custom Sequencing Primers					
Search for NGS Services	(methylation) and SNP identification are best served by paired-end rur costly and time consuming, you get back twice the amount of data at I sequence	ns. While paired end runs are more less than double the cost to			

Minimizing Bias

Generally try to use a minimal number of PCR cycles during library prep.

Use high-fidelity polymerases that exhibit a low amplification bias (KAPA HiFi or NEB Q5)

Discussion of additional sources of bias: van Dijk et al. 201 https://www.ncbi.nlm.nih.gov/pubmed/24440557

